REVA UNIVERSITY
Bengaluru, India

SCHOOL OF APPLIED SCIENCES

M.Sc., (Microbial Technology) Program

Hand Book

2021-2023.

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Chancellor's Message

"Education is the most powerful weapon which you can use to change the world."

- Nelson Mandela.

There was a time when survival depended on just the realization of physiological needs. We are indeed privileged to exist in a time when 'intellectual gratification' has become indispensable. Information is easily attainable for the soul that is curious enough to go look for it. Technological boons enable information availability anywhere anytime. The difference, however, lies between those who look for information and those who look for knowledge.

It is deemed virtuous to serve seekers of knowledge and as educators it is in the ethos at REVA University to empower every learner who chooses to enter our portals. Driven by our founding philosophy of 'Knowledge is Power', we believe in building a community of perpetual learners by enabling them to look beyond their abilities and achieve what they assumed impossible.

India has always been beheld as a brewing pot of unbelievable talent, acute intellect and immense potential. All it takes to turn those qualities into power is a spark of opportunity. Being at a University is an exciting and rewarding experience with opportunities to nurture abilities, challenge cognizance and gain competence.

For any University, the structure of excellence lies in the transitional abilities of its faculty and its facility. I'm always in awe of the efforts that our academic board puts in to develop the team of subject matter experts at REVA. My faculty colleagues understand our core vision of empowering our future generation to be ethically, morally and intellectually elite. They practice the art of teaching with a student-centred and transformational approach. The excellent infrastructure at the University, both educational and extra-curricular, magnificently demonstrates the importance of ambience in facilitating focused learning for our students.

A famous British politician and author from the 19th century - Benjamin Disraeli, once said 'A University should be a place of light, of liberty and of learning'. Centuries later this dictum still inspires me and I believe, it takes team-work to build successful institutions. I welcome you to REVA University to join hands in laying the foundation of your future with values, wisdom and knowledge.

Dr. P. Shyama Raju
The Founder and Hon'ble Chancellor, REVA University

Vice-Chancellor's Message

The last two decades have seen a remarkable growth in higher education in India and across the globe. The move towards inter-disciplinary studies and interactive learning have opened up several options as well as created multiple challenges. India is at a juncture where a huge population of young crowd is opting for higher education. With the tremendous growth of privatization of education in India, the major focus is on creating a platform for quality in knowledge enhancement and bridging the gap between academia and industry.

A strong believer and practitioner of the dictum "Knowledge is Power", REVA University has been on the path of delivering quality education by developing the young human resources on the foundation of ethical and moral values, while boosting their leadership qualities, research culture and innovative skills. Built on a sprawling 45 acres of green campus, this 'temple of learning' has excellent and state-of-the-art infrastructure facilities conducive to higher teaching-learning environment and research. The main objective of the University is to provide higher education of global standards and hence, all the programs are designed to meet international standards. Highly experienced and qualified faculty members, continuously engaged in the maintenance and enhancement of student-centric learning environment through innovative pedagogy, form the backbone of the University.

All the programs offered by REVA University follow the Choice Based Credit System (CBCS) with Outcome Based Approach. The flexibility in the curriculum has been designed with industry-specific goals in mind and the educator enjoys complete freedom to appropriate the syllabus by incorporating the latest knowledge and stimulating the creative minds of the students. Bench marked with the course of studies of various institutions of repute, our curriculum is extremely contemporary and is a culmination of efforts of great think-tanks - a large number of faculty members, experts from industries and research level organizations. The evaluation mechanism employs continuous assessment with grade point averages. We believe sincerely that it will meet the aspirations of all stakeholders – students, parents and the employers of the graduates and postgraduates of REVA University.

At REVA University, research, consultancy and innovation are regarded as our pillars of success. Most of the faculty members of the University are involved in research by attracting funded projects from various research level organizations like DST, VGST, DBT, DRDO, AICTE and industries. The outcome of the research is passed on to students through live projects from industries. The entrepreneurial zeal of the students is encouraged and nurtured through EDPs and EACs.

REVA University has entered into collaboration with many prominent industries to bridge the gap between industry and University. Regular visits to industries and mandatory internship with industries have helped our students. REVA University has entered into collaboration with many prominent industries to bridge the gap between industry and University. Regular visits to industries and mandatory internship with industries have helped our students become skilled with relevant to industry requirements. Structured training programs on soft-skills and preparatory training for competitive exams are offered here to make students more employable. 100% placement of eligible students speaks the effectiveness of these programs. The entrepreneurship development activities and establishment of "Technology Incubation Centres" in the University extend full support to the budding entrepreneurs to nurture their ideas and establish an enterprise.

With firm faith in the saying, "Intelligence plus character –that is the goal of education" (Martin Luther King, Jr.), I strongly believe REVA University is marching ahead in the right direction, providing a holistic education to the future generation and playing a positive role in nation building. We reiterate our endeavour to provide premium quality education accessible to all and an environment for the growth of over-all personality development leading to generating "GLOBAL PROFESSIONALS".

Welcome to the portals of REVA University!

Dr. M. Dhanamjaya
Vice Chancellor

Director or Dean's Message

Microbial Technology as interdisciplinary subject assimilates in itself a number of disciplines and as such has grown rapidly. M Sc in Microbial Technology offered by REVA University aims to provide the required skills and knowledge necessary to pursue a successful career in Microbiology. This program imparts need based, practical education in contemporary world to develop global competence among students. It strives to prepare students to become leaders in the field of Life Sciences in general and Microbiology in particular by encouraging them to inculcate scientific thinking coupled with creative and innovative ideas.

MSc Microbial Technology provides the specialization of three major domains of microbiology such as Industrial & Food Microbiology, Agriculture & Environmental Microbiology and Medical & Diagnostic Microbiology based on the interest of the students in collaboration with the industry partners. The program provides hands-on and industry based training and practical skills in the specialised field of Microbiology and basics of Molecular genetics, Bioinformatics, Biochemical techniques & Enzymology, Environmental Microbiology, Molecular biology and Applied Microbiology aligning to current demand in the field of research & industry.

As far as employment is concerned Microbiology has become one of the fast-growing sectors. Employment record shows that Microbiology has a great scope in future. Microbiologists can find careers with pharmaceutical companies, chemical, agricultural and allied companies. They can be employed in the areas of planning, production and management of bio-processing industries. There is a large-scale employment in research laboratories run by the government as well as the corporate sectors. Further, there is great demand for biotechnology experts in numerous industries and sectors after the completion of MSc Biotechnology course, some of which are: Agriculture, Animal Husbandry, Environment Conservation, Genetic Engineering, Health Care, Medicine, Industrial Research and Development.

This handbook provides you outline of regulations for master's degree, scheme of instruction, and detailed syllabus. I am sure the students choosing MSc Microbiology at REVA University will enjoy the curriculum, teaching and learning environment, the vast infrastructure and the experienced faculties with excellent involvement and guidance. We will strive to provide all needed comfort and congenial environment for their studies. I wish all students a pleasant stay at REVA and grand success in their career.

Prof. Shilpa B R
Deputy Director
School of Applied Sciences

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RUKMINI EDUCATIONAL CHARITABLE TRUST

It was the dream of late Smt. Rukmini Shyama Raju to impart education to millions of underprivileged children as she knew the importance of education in the contemporary society. The dream of Smt. Rukmini Shyama Raju came true with the establishment of Rukmini Educational Charitable Trust (RECT), in the year 2002. Rukmini Educational Charitable Trust (RECT) is a Public Charitable Trust, set up in 2002 with the objective of promoting, establishing and conducting academic activities in the fields of Arts, Architecture, Commerce, Education, Engineering, Environmental Science, Legal Studies, Management and Science & Technology, among others. In furtherance of these objectives, the Trust has set up the REVA Group of Educational Institutions comprising of REVA Institute of Technology & Management (RITM), REVA Institute of Science and Management (RISM), REVA Institute of Management Studies (RIMS), REVA Institute of Education (RIE), REVA First Grade College (RFGC), REVA Independent PU College at Kattigenahalli, Ganganagar and Sanjaynagar and now REVA University. Through these institutions, the Trust seeks to fulfil its vision of providing world class education and create abundant opportunities for the youth of this nation to excel in the areas of Arts, Architecture, Commerce, Education, Engineering, Legal Studies, Management and Science & Technology.

Every great human enterprise is powered by the vision of one or more extraordinary individuals and is sustained by the people who derive their motivation from the founders. The Chairman of the Trust is Dr. P. Shyama Raju, a developer and builder of repute, a captain of the industry in his own right and the Chairman and Managing Director of the DivyaSree Group of companies. The idea of creating these top notched educational institutions was born of the philanthropic instincts of Dr. P. Shyama Raju to do public good, quite in keeping with his support to other socially relevant charities such as maintaining the Richmond road park, building and donating a police station, gifting assets to organizations providing accident and trauma care, to name a few.

The Rukmini Educational Charitable Trust drives with the main aim to help students who are in pursuit of quality education for life. REVA is today a family of ten institutions providing education from PU to Post Graduation and Research leading to PhD degrees. REVA has well qualified experienced teaching faculty of whom majority are doctorates supported by committed administrative and technical staff. Over 15,000+ students study various courses across REVA's three campuses equipped with exemplary state-of-the-art infrastructure and conducive environment for the knowledge driven community.

ABOUT REVA UNIVERSITY

REVA University has been established under the REVA University Act, 2012 of Government of Karnataka and notified in Karnataka State Gazette No. 80 dated 27thFebruary, 2013. The University is empowered by UGC to award degrees any branch of knowledge under Sec.22 of the UGC Act. The University is a Member of Association of Indian Universities, New Delhi. The main objective of the University is to prepare students with knowledge, wisdom and patriotism to face the global challenges and become the top leaders of the country and the globe in different fields.

REVA University located in between Kempegowda International Airport and Bangalore city, has a sprawling green campus spread over 45 acres of land and equipped with state-of-the-art infrastructure that provide conducive environment for higher learning and research. The REVA campus has well equipped laboratories, custom-built teaching facilities, fully air-conditioned library and central computer centre, the well-planned sports facility with cricket ground, running track & variety of indoor and outdoor sports activities, facilities for cultural programs. The unique feature of REVA campus is the largest residential facility for students, faculty members and supportive staff.

REVA consistently ranked as one of the top universities in various categories because of the diverse community of international students and its teaching excellence in both theoretical and technical education in the fields of Engineering, Management, Law, Science, Commerce, Arts, Performing Arts, and Research Studies. REVA offers 28 Undergraduate Programmes, 22 Full-time and 2 Part-time Postgraduate Programmes, 18 Ph. D Programmes, and other Certificate/ Diploma/Postgraduate Diploma Programmes in various disciplines.

The curriculum of each programme is designed with a keen eye for detail by giving emphasis on hands-on training, industry relevance, social significance, and practical applications. The University offers world-class facilities and education that meets global standards.

The programs being offered by the REVA University are well planned and designed after detailed study with emphasis with knowledge assimilation, applications, global job market and their social relevance. Highly qualified, experienced faculty and scholars from reputed universities / institutions, experts from industries and business sectors have contributed in preparing the

scheme of instruction and detailed curricula for this program. Greater emphasis on practice in respective areas and skill development to suit to respective job environment has been given while designing the curricula. The Choice Based Credit System and Continuous Assessment Graded Pattern (CBCS – CAGP) of education has been introduced in all programs to facilitate students to opt for subjects of their choice in addition to the core subjects of the study and prepare them with needed skills. The system also allows students to move forward under the fast track for those who have the capabilities to surpass others. These programs are taught by well experienced qualified faculty supported by the experts from industries, business sectors and such other organizations. REVA University has also initiated many supportive measures such as bridge courses, special coaching, remedial classes, etc., for slow learners so as to give them the needed input and build in them confidence and courage to move forward and accomplish success in their career. The University has also entered into MOUs with many industries, business firms and other institutions seeking their help in imparting quality education through practice, internship and also assisting students' placements.

REVA University recognizing the fact that research, development and innovation are the important functions of any university has established an independent Research and Innovation division headed by a senior professor as Dean of Research and Innovation. This division facilitates all faculty members and research scholars to undertake innovative research projects in engineering, science & technology and other areas of study. The interdisciplinary-multidisciplinary research is given the top most priority. The division continuously liaisons between various funding agencies, R&D Institutions, Industries and faculty members of REVA University to facilitate undertaking innovative projects. It encourages student research projects by forming different research groups under the guidance of senior faculty members. Some of the core areas of research wherein our young faculty members are working include Data Mining, Cloud Computing, Image Processing, Network Security, VLSI and Embedded Systems, Wireless Censor Networks, Computer Networks, IOT, MEMS, Nano- Electronics, Wireless Communications, Bio-fuels, Nano-technology for coatings, Composites, Vibration Energies, Electric Vehicles, Multilevel Inverter Application, Battery Management System, LED Lightings, Renewable Energy Sources and Active Filter, Innovative Concrete Reinforcement, Electro Chemical Synthesis, Energy Conversion Devices, Nano-structural Materials, Photo-electrochemical Hydrogen generation, Pesticide Residue Analysis, Nano materials, Photonics, Nana Tribology, Fuel Mechanics, Operation Research, Graph theory, Strategic Leadership and Innovative Entrepreneurship, Functional Development Management, Resource Management and

Sustainable Development, Cyber Security, General Studies, Feminism, Computer Assisted Language Teaching, Culture Studies etc.

The REVA University has also given utmost importance to develop the much required skills through variety of training programs, industrial practice, case studies and such other activities that induce the said skills among all students. A full-fledged Career Development and Placement (CDC) department with world class infrastructure, headed by a dynamic experienced Professor& Dean, and supported by well experienced Trainers, Counsellors and Placement Officers.

The University also has University-Industry Interaction and Skill Development Centre headed by a Senior Professor & Director facilitating skill related training to REVA students and other unemployed students. The University has been recognised as a Centre of Skill Development and Training by NSDC (National Skill Development Corporation) under Pradhan Mantri Kaushal VikasYojana. The Centre conducts several add-on courses in challenging areas of development. It is always active in facilitating student's variety of Skill Development Training programs.

The University has collaborations with Industries, universities abroad, research institutions, corporate training organizations, and Government agencies such as Florida International University, Oklahoma State University, Western Connecticut University, University of Alabama, Huntsville, Oracle India Ltd, Texas Instruments, Nokia University Relations, EMC^2, VMware, SAP, Apollo etc, to facilitate student exchange and teacher–scholar exchange programs and conduct training programs. These collaborations with foreign universities also facilitates students to study some of the programs partly in REVA University and partly in foreign university, viz, M.S in Computer Science one year in REVA University and the next year in the University of Alabama, Huntsville, USA.

The University has also given greater importance to quality in education, research, administration and all activities of the university. Therefore, it has established an independent Internal Quality division headed by a senior professor as Dean of Internal Quality. The division works on planning, designing and developing different quality tools, implementing them and monitoring the implementation of these quality tools. It concentrates on training entire faculty to adopt the new tools and implement their use. The division further works on introducing various examination and administrative reforms.

To motivate the youth and transform them to become innovative entrepreneurs, successful leaders of tomorrow and committed citizens of the country, REVA organizes interaction between students and successful industrialists, entrepreneurs, scientists and such others from time to time. As a part of this exercise great personalities such as Bharat Ratna Prof. C. N. R. Rao, a renowned Scientist, Dr. N R Narayana Murthy, Founder and Chairman and Mentor of Infosys, Dr. K Kasturirangan, Former Chairman ISRO, Member of Planning Commission, Government of India, Dr.Balaram, Former Director IISc., and noted Scientist, Dr. V S Ramamurthy, Former Secretary, DST, Government of India, Dr. V K Aatre, noted Scientist and former head of the DRDO and Scientific Advisor to the Ministry of Defence Dr.Sathish Reddy, Scientific Advisor, Ministry of Defence, New Delhi and many others have accepted our invitation and blessed our students and faculty members by their inspiring addresses and interaction.

REVA organises various cultural programs to promote culture, tradition, ethical and moral values to our students. During such cultural events the students are given opportunities to unfold their hidden talents and motivate them to contribute innovative ideas for the progress of the society. The event not only gives opportunities to students of REVA but also students of other Universities and Colleges. During three days of this mega event students participate in debates, Quizzes, Group discussion, Seminars, exhibitions and variety of cultural events. Another important event is Shubha Vidaaya, Graduation Day for the final year students of all the programs, wherein, the outgoing students are felicitated and are addressed by eminent personalities to take their future career in a right spirit, to be the good citizens and dedicate themselves to serve the society and make a mark in their respective spheres of activities. During this occasion, the students who have achieved top ranks and won medals and prizes in academic, cultural and sports activities are also recognised by distributing awards and prizes. The founders have also instituted medals and prizes for sports achievers every year. The physical education department conducts regular yoga class's every day to students, faculty members, administrative staff and their family members and organizes yoga camps for villagers around.

VISION

REVA University aspires to become an innovative university by developing excellent human resources with leadership qualities, ethical and moral values, research culture and innovative skills through higher education of global standards

MISSION

- To create excellent infrastructure facilities and state-of-the-art laboratories and incubation centres
- To provide student-centric learning environment through innovative pedagogy and education reforms
- To encourage research and entrepreneurship through collaborations and extension activities
- To promote industry-institute partnerships and share knowledge for innovation and development
- To organize society development programs for knowledge enhancement in thrust areas
- To enhance leadership qualities among the youth and enrich personality traits, promote patriotism and moral values.

OBJECTIVES

- Creation, preservation and dissemination of knowledge and attainment of excellence in different disciplines
- Smooth transition from teacher - centric focus to learner - centric processes and activities
- Performing all the functions of interest to its major constituents like faculty, staff, students and the society to reach leadership position
- Developing a sense of ethics in the University and Community, making it conscious of its obligations to the society and the nation
- Accepting the challenges of globalization to offer high quality education and other services in a competitive manner

ABOUT THE SCHOOL OF APPLIED SCIENCES

The School of Applied Sciences offers graduate and post graduate programs in Biotechnology, Biochemistry, Chemistry, Physics and Mathematics which are incredibly fascinating. It aims to attract talented youth and train them to acquire knowledge and skills useful to industrial sectors, research laboratories, and educational institutions. The School presently offers M.Sc. degree programs in Bio-Chemistry, Bio-Technology, Chemistry, Physics, Mathematics, Bioinformatics, Microbial Technology and B Sc with various combinations viz, Biotechnology, Microbiology, Biochemistry and Genetics, Medical Laboratory Technology, Physics, Chemistry, Mathematics, Statistics, Computer Science, Bioinformatics, Statistics & Computer Science, Microbiology, Chemistry and Genetics and also Post Graduate Diploma in Clinical Embryology and Artificial Reproductive Technology. The school also facilitates research leading to PhD in Biotechnology, Biochemistry, Physics, Chemistry, Mathematics and related areas of study.

The School of Applied Sciences is shouldered by well qualified, experienced and highly committed faculty. The state-of-the-art infrastructure digital classrooms, well equipped laboratories, conference rooms and the serene academic atmosphere at REVA University will enhance the transfer as well as creation of knowledge. The school provides an interactive, collaborative peer tutoring environment that encourages students to break down complex problems and develop strategies for finding solutions across a variety of situations and disciplines. The school aims to develop a learning community of critical thinkers who serves as models of innovative problems solving in the university environment to enrich their academic and professional careers.

VISION

To nurture intellect, creativity, character, professionalism and research culture among students and impart contemporary knowledge in various branches of Chemical, Biological, Physical and Mathematical Sciences that are socially relevant and transform them to become global citizens with leadership qualities.

MISSION

1. To achieve excellence in studies and research through pedagogy and support interface between industry and academia
2. To create intellectual curiosity, academic excellence, and integrity through multidimensional exposure
3. To establish state of the art laboratories to support research and innovation and promote mastery of science.
4. To inculcate an ethical attitude and make students competitive to serve the society and nation.

BOS MEMBERS

Sl. No	Name, Designation &Affiliation	External/Internal member
1	**Prof. Shilpa B.R** Associate Professor, Deputy Director and Head of the Department, Department of Biotechnology, School of Applied Sciences, REVA University, Bengaluru, Karnataka, India.	Chairperson
2	**Dr. Mahesh M,** **Chief Executive Officer**, Azyme Biosciences Pvt. Ltd., Bengaluru, Karnataka, India	Industry Member
3	**Dr. Basavaraj Girennavar** Chairman and Managing Director, Criyagen Agri & Biotech Pvt. Ltd., Bengaluru, Karnataka, India	Industry Member
4	**Dr. Latha P,** Chief Operating Officer, X-Cyton Pvt Ltd, Bengaluru, Karnataka, India	Industry Member
5	**Dr. Pannuru Padmavathi,** Managing Director, DR Biosciences LLP, Bengaluru, Karnataka, India	Industry Member
6	**Dr. Pasupuleti Visweswara Rao** Associate Professor & Research Coordinator Head of Unit-Non-Communicable Diseases Research Niche Areas, Head of Internationalization unit Department of Biomedical Sciences and Therapeutics Universiti Malaysia Sabah (Central University) Kota Kinabalu, Sabah, Malaysia	International and Alumni Member
7	**Dr. Mahesh Yandigeri,**	Academic Member

	Senior Scientist, Division of Genomic Resources, National Bureau of Agricultural Insect Resources (NBAIR), Hebbal, Bengaluru, Karnataka, India	
8	**Dr. G.B. Manjunatha Reddy,** Scientist, ICAR - National Institute of Veterinary Epidemiology and Disease Informatics (ICAR – NIVEDI), Yelahanka, Bangalore, Karnataka, India	Academic Member
8	**Dr. V. Veena** Assistant Professor Department of Biotechnology School of Applied Sciences REVA University, Bangalore, Karnataka, India	Internal Member
9	**Dr. R. Ramachandra** Assistant Professor Department of Biotechnology School of Applied Sciences REVA University, Bangalore, Karnataka, India	Internal Member
10	**Dr. N. M. Guruprasad** Assistant Professor Department of Biotechnology School of Applied Sciences REVA University, Bangalore, Karnataka, India	Internal Member

M.Sc. Microbial Technology Program Overview

The Masters' program in **Microbial Technology is Industry Oriented Program**, majorly focused on the applied aspects of microbiology in various specialized areas of science and technology such as industry, agriculture and medicine. Microbial Technology emphasizes hands-on learning through laboratories sessions, internships, skill development programs conducted every semester.

The curriculum of the program is reviewed and approved esteemed members from reputed industries and research organization to makes is industry oriented in true sense. Students will have an opportunity to take up projects either at department or at reputed industries as well at state and central governments research organizations in the final semester. Well qualified faculty members with rich industry and research experience guide the students to excel in academics and research. Three different specializations have been offered for the program like Industrial and Food Microbiology, Plant and Agricultural Microbiology and Medical and Pharmaceutical Microbiology in collaboration with reputed industries like **Azyme Biosciences Pvt. Ltd., Criyagen Agri & Biotech Pvt. Ltd., DR BioSciences LLP., X-Cyton Pvt Ltd.,** to ensure right skill sets for students as per chosen specialization.

Curriculum is well designed with hardcore (HC) and softcore (SC) courses. MOOC/Swayam/ Coursera Online Courses, short term certification courses and workshops are offered to equip the students with skill sets required for employability and research. There are wide range of career options for MSc Microbial Technology program in various domains of life sciences. The career opportunities include teaching, Scientist, Industries, Research Organizations, State and Central Universities, Agriculture Universities, Hospitals, Diagnostic laboratories, Nutritionist, Project Assistant, Research Assistant, Research Analyst, Clinical Research Associates, Teaching Assistant, Lecturer and Business start-ups. In this context, University Programmes at undergraduate and postgraduate level in Life Sciences across the country have become relevant.

M. Sc. Microbial Technology at REVA UNIVERSITY has been designed to meet the human resources needs of existing and futuristic Life Science industries, research organizations and academic institutions. The programme is designed to produce graduates with higher order critical,

analytical, problem solving and research skills, ability to think rigorously and independently to meet higher level expectations of biotech industries, research organization and academic institutions. The programme also provides sufficient skills and training on entrepreneurship development in Life Sciences.

Program Educational Objectives (PEOs)

After few years of graduation, the graduates will:

PEO-1	Become a professional Microbiologist with academic excellence, ethics and communication skills and perform in a team.
PEO-2	Equip with necessary analytical and research skills to establish themselves as a competent scientist/microbiologist.
PEO-3	Become successful entrepreneurs with an ability to develop new products/processes with an attitude of lifelong learning.

Program Outcomes (POs)

After successful completion of program, the graduates will be able

1. **Master of Science Knowledge**-Apply the knowledge of microbiology and applied microbiology to the solution of complex biological problems.
2. **Problem analysis**- Identify, formulate, review research literature and analyse complex biological problems reaching substantiated conclusions using various principles of life science domain and microbiology.
3. **Design/development of solutions**-Design solutions for complex biological problems and design protocols or processes that meet the specified needs with appropriate consideration for the public health and safety, conservation of biodiversity, better understanding of the microorganisms and necessary tools for finding solutions of various crippling human/plant diseases with ethical, societal, and environmental considerations.
4. **Conduct investigations of complex problems**-Use the various protocols developed through extensive research-based knowledge and methods including design of experiments, analysis and interpretation of data, and provide valid and reproducible conclusions.
5. **Environment and sustainability**-Apply the classic and modern biological theoretical and practical knowledge gained to address societal, health, microbial and plant biodiversity

studies, safety, ethical and cultural issues and the consequent responsibilities relevant to the professional upgradation of the student and society as a need for sustainable development.

6. **Ethics-**Apply ethical principles established by different government agencies and commit to research ethics, responsibilities and norms to undertake their current and future research and development.
7. **Individual and team work**-Be an independent thinker and researcher effectively as an individual, and as a member or leader of different teams, and in multidisciplinary research Institutions and Universities.
8. **Communication**-Communicate effectively on complex research activities with the scientific community and with society at large, as a scientist or a teacher, be well versed with scientific writing and write effective reports and design research projects, make effective presentations, and be able to defend it efficiently.
9. **Project management and finance**-Write good research and development projects relevant to the needs of society and environment and attract extra mural funds for himself and his team in the Institute or University from various funding agencies and manage R&D projects effectively.
10. **Life-long learning**-Apply the discipline, ethics and knowledge obtained to engage in independent and life-long learning in their respective fields of interest wherever they go for further higher studies or jobs.

Program Specific Outcomes (PSO)

After successful completion of the programme, the graduates shall be able to

1. Understand about environmental microbiology, food & diary Microbiology, Industrial Microbiology and many other technologies involved in microbiology industries and academic institutions.
2. Provide a strong foundation for a career working skills with project management, business development or venture capital within the microbiology, agriculture, food, environment, medical technology or related industries.
3. Utilise the skills to design, perform and analyse a research problem which has a social relevance and outcome with acquired skills of presentation and scientific writing.

REVA University Regulations for Choice Based Credit System (CBCS) and Continuous Assessment Grading Pattern (CAGP) for Postgraduate Degree programs- 2021-22

(Framed as per the provisions under Section 35 (ii), Section 7 (x) and Section 8 (xvi) & (xxi) of the REVA University Act, 2012)

1. Tie and Commencement:

1.1. These Regulations shall be called the **"REVA University Regulations for Choice Based Credit System (CBCS) and Continuous Assessment Grading Pattern (CAGP) for Post Graduate Degree Programs- 2021-22".**

1.2. These Regulations shall come into force from the date of assent of the Chancellor.

2. The Programs:

The following programs and all Graduate Degree programs to be instituted and introduced in REVA University in coming years shall follow these regulations.

M.Sc. in:

Biotechnology
Biochemistry
Chemistry
Physics
Mathematics
Microbial Technology
Bioinformatics

3. Definitions:

Course: Every course offered will have three components associated with the teaching-learning process of the course, namely:

(i) L= Lecture (ii) T= Tutorial (iii) P=Practice; where:

L stands for **Lecture** session consisting of classroom instruction.

T stands for **Tutorial** session consisting participatory discussion / self-study/ desk work/ brief seminar presentations by students and such other novel methods that make a student to absorb and assimilate more effectively the contents delivered in the Lecture classes.

P stands for **Practice** session and it consists of Hands on Experience / Laboratory Experiments / Field Studies / Case Studies that equip students to acquire the much required skill component.

4. Courses of study and Credits

4.1. The study of various subjects in M. Sc., degree program is grouped under various courses. Each of these course carries credits which are based on the number of hours of teaching and learning.

4.1.1. In terms of credits, every **one hour session of L amounts to 1 credit per Semester.**
In terms of credits, every **one hour session of L amounts to 1 credit per Semester** and a minimum of **two hour session of T or P amounts to 1 credit per Semester** over a period of one Semester of 16 weeks for teaching-learning process.

4.1.2. The total duration of a semester is 20 weeks inclusive of semester-end examination.

4.1.3. A course shall have either or all the four components. That means a course may have only lecture component, or only practical component or combination of any two or all the three components.

4.1.4. ***The concerned BoS will assign Credit Pattern for every course based on the requirement. However, generally, courses can be assigned with 1-4 Credits depending on the size of the course.***

4.1.5. Different **Courses of Study** are labelled and defined as follows:

Core Course:
A course which should compulsorily be studied by a candidate as a core-requirement is termed as a Core course. The CORE courses of Study are of THREE types, viz – (i) Hard Core Course, and (ii) Soft Core Course, (iii) Open Elective and (iV) Mandatory Course (MC)

a. Hard Core Course (HC):

The **Hard Core Course** is a Core Course in the main branch of study and related branch(es) of study, if any that the candidates have to complete compulsorily.

b. Soft Core Course (SC):

A Core course may be a **Soft Core** if there is a choice or an option for the candidate to choose a course from a pool of courses from the main branch of study or from a sister/related branch of study which supports the main branch of study.

c. Open Elective Course:

An elective course chosen generally from other discipline / related subject, with an intention to seek exposure to the related subjects other than the main discipline the student is studying is called an O**pen Elective Course.**

d. Mandatory Course (MC): The mandatory course is non credited but must be passed in order to complete the Graduate Degree Program

d. Skill Enhancement Course:

It is a mandatory course to equip students with skill sets required as per the industry expectation. Candidate will seek exposure through workshops and other certificate-based courses.

e. Project Work / Dissertation:

Project work / Dissertation work is a special course involving application of knowledge in solving / analysing /exploring a real life situation / difficult problem. A project work carrying **FOUR or SIX** credits is called **Minor Project** work / **Dissertation.** A project work of **EIGHT, TEN, TWELVE or SIXTEEN** credits is called **Major Project** work / **Dissertation**. **A Project work may be a hard core or a Soft Core as decided by the BoS / concerned.**

5. Eligibility for Admission:

Bachelors Degree of three years with Biotechnology or any Life Science subject as one of the cognate / major / optional subjects with 60% (40% in case of candidates belonging to SC/ST) of marks in aggregate from any recognized University / Institution or any other qualification recognized as equivalent thereto.

6. Scheme, Duration and Medium of Instructions:

6.1. M.Sc., degree program is of 4 semesters - 2 years duration. A candidate can avail a maximum of 6 semesters (3 years) including blank semesters, if any to successfully complete M.Sc. degree. Whenever a candidate opts for blank semester, he/she has to study the prevailing courses offered by the School when he/she resumes his/her studies.

6.2. The medium of instruction shall be English.

7. Credits and Credit Distribution

7.1. A candidate has to earn 90 credits for successful completion of Two-Year Postgraduate degree with a distribution of credits for different courses as given in Table - 1 given below:

Table-1: Credits and Credit Distribution for Two Year Post Graduate degree program in Sciences

Course Type	**Credits for Two Year (4 Semesters) Post Graduate Degree Programs**
Hard Core Course	A minimum of 60 but not exceeding 70
Soft Core Course	A minimum of 10 but not exceeding 30
Open Elective	A minimum of 4
SEC	A minimum of 2
Project/Dissertation	A minimum of 10
MOOC / Swayam/ Coursera/Soft Skill Training	A minimum of 4
Total	**90**

7.2. The concerned BOS based on the credits distribution pattern given above shall prescribe the credits to various types of courses and shall assign title to every course including project work, practical work, field work, self-study elective, as **Hard Core (HC) or Soft Core (SC) Open Elective (OE)**, **Skill Enhancement course (SEC)** and **Mandatory Course (MC)**

7.3. Every course including project work, practical work, field work, self-study elective should be entitled as Hard Core (HC) or Soft Core (SC) or Open Elective (OE), SEC (Skill Enhance Course), and Mandatory Course (MC) by the BoS concerned.

The BOS of respective program/ discipline shall decide about the total credits for the courses.

Sl. No.	Course Title	Number of Credits
1	MOOC / Swayam/ Coursera/Internship /Soft Skill Training	4
	Total	**4**

7.4. The concerned BOS shall specify the desired Program Objectives, Program Educational Objectives, Program Specific Outcomes and Course Outcomes while preparing the curriculum of a particular program.

7.5. A candidate can enrol for a maximum of 30 credits and a minimum of 20 credits per Semester. However, he / she may not successfully earn a maximum of 30 credits per semester. This maximum of 30 credits does not include the credits of courses carried forward by a candidate.

7.6. Only such full-time candidates who register for a minimum prescribed number of credits in each semester from I semester to IV semester and complete successfully 90 credits in 4 successive semesters shall be considered for declaration of Ranks, Medals, Prizes and are eligible to apply for Student Fellowship, Scholarship, Free ships, and such other rewards / advantages which could be applicable for all full-time students and for hostel facilities.

8. Add-on Proficiency Certification / Diploma:

8.1 Add- on Proficiency Certification:

To acquire Add on Proficiency Certification a candidate can opt to complete a minimum of 4 extra credits either in the same discipline /subject or in different discipline / subject in excess to 90 credits for the Two Year Post Graduate degree programs.

8.2 Add on Proficiency Diploma:

To acquire Add on Proficiency Diploma, a candidate can opt to complete a minimum of 18 extra credits either in the same discipline /subject or in different discipline / subject in excess to 90 credits for the Two Year Post Graduate degree programs.

The Add on Proficiency Certification / Diploma so issued to the candidate contains the courses studied and grades earned.

9. Assessment and Evaluation

a) Each course is assessed for a total weight of 100%. Out of the total 100% weight; 50% weight is for Continuous Internal Assessment (CIA or IA) and the remaining 50% for the Semester End Examination (SEE). This applicable for theory, laboratory, workshop, studio and any such courses

b) Out of 50% weight earmarked for Internal Assessment (IA)- 15% for test-1, 15% for test-2 and 20% for Assignments and this is applicable for theory based courses

c) The tests and assignments are conducted as per the semester academic calendar provided by the University

The details as given in the table

Component	**Description**	**Conduction**	**Weight Percentage**
C1	**Test-1: IA1**	**8th week from the starting date of semester**	**15**
	Test-2: IA2	**16th week from the starting date of semester**	**15**
C2	**1 Assignment**	**7th week**	**10**
	2 Assignment	**14th week**	**10**
C3	**SEE including practical**	**between 17th Week-20th Week**	**50**
Results to be Announced			**By the end of 21st Week**

Note: IA or CIA includes C1 and C2

Each test must be conducted for a duration of 60 minutes, setting the test question paper for a maximum of 30 marks. The final examination must be conducted for a duration of 3 hours and the question paper must be set for a maximum of 100 marks.

d) Students are required to complete courses like technical skills, placement related courses, Open electives and any such value addition or specialized courses through online platforms like SWAYAM/NPTEL/Any other reputed online education aggregator. Students are required to choose the courses on the advice of their course coordinator/Director and required to submit the course completion certificate along with percentage of marks/grade scored in the assessment conducted by the online education aggregator. If the online education aggregator has issued a certificate along with the grade or marks scored to students, such courses will be considered for SGPA calculations, in case the aggregator has issued only a certificate and not marks scored, then such courses will be graded through an examination by concerned School, in case, if grading is not possible, students will be given a pass grade and award the credit and the credits will not be considered for SGPA calculations. The Online/MOOCs courses will not have continuous internal assessment component

Such of those students who would like to discontinue with the open elective course that they have already registered for earning required credits can do so, however, they need to complete the required credits by choosing an alternative open elective course.

Setting question paper and evaluation of answer scripts.

1. For SEE, three sets of question papers shall be set for each theory course out of which two sets will be by the internal examiners and one set will be by an external examiner. In subsequent years by carrying forward the unused question papers, an overall three sets of question papers should be managed and depending on the consumption of question papers either internal or external examiner be called for setting the question paper to maintain an overall tally of 3 papers with the conditioned mentioned earlier. The internal examiner who sets the question paper should have been course tutor.
2. The Chairman of BoE shall get the question papers set by internal and external examiners.
3. The Board of Examiners shall scrutinize and approve the question papers and scheme of valuation. It is the responsibility of the BoE to see that all questions contained in the question paper are within the prescribed syllabus of the concerned course.
4. There shall be single valuation for all theory papers by internal examiners. However, there shall be moderation by the external examiner who has the subject background. In case no external examiner with subject background is available, a senior faculty member within the discipline shall be appointed as moderator.
5. The SEE examination for Practical work / Field work / Project work/Internship will be

conducted jointly by internal and external examiners as detailed below: However, the BoE on its discretion can also permit two internal examiners.

6. If a course is fully of (L=0):T:(P=0) type or a course is partly P type i.e, (L=3): (T=0) (P=1), then the examination for SEE component will be as decided by the BoS concerned.

10. Evaluation of Practical's and Minor Project / Major Project / Dissertation

10.3.1. A practical examination shall be assessed on the basis of:

a) Knowledge of relevant processes;

b) Skills and operations involved;

c) Results / products including calculation and reporting.

10.3.2. In case a course is fully of P type (L=0:T=0:P=4), 2 credits, the performance of a candidate shall be assessed for a maximum of 100 marks as explained below:

a) Continuous Internal assessment (CIA) = 50 marks

b) Semester end practical examination (SEE) = 50 marks

The 50 marks for continuous assessment shall further be allocated as under (IA or CIA):

i	Conduction of regular practical throughout the semester	20 marks
ii	Maintenance of lab records /industry reports/SDP reports	15 marks
iii	Laboratory test and viva	15 marks
	Total	50 marks

The 50 marks meant for Semester End Examination, shall be allocated as under:

i	Conduction of semester end practical examination	30 marks
ii	Write up about the experiment / practical conducted	10 marks
iii	Viva Voce	10 marks
	Total	50 marks

10.3.3. The SEE for Practical work will be conducted jointly by internal and external examiners. However, if external examiner does not turn up, then both the examiners will be internal examiners.

10.3.4. In case a course is partly P type i.e, (L=3): (T=0) (P=1), then the examination for SEE

component will be as decided by the BoS concerned.

10.3.5. The duration for semester-end practical examination shall be decided by the concerned School Board.

10.4. Evaluation of Minor Project / Major Project / Dissertation:

Right from the initial stage of defining the problem, the candidate has to submit the progress reports periodically and also present his/her progress in the form of seminars in addition to the regular discussion with the supervisor. At the end of the semester, the candidate has to submit final report of the project / dissertation, as the case may be, for final evaluation. The components of evaluation are as follows:

1	First Dissertation presentation describing the problem definition	Should be done a semester before the project semester	Weightage: 0%
2	Dissertation Progress presentation-1	7th week from the start date of project semester	Weightage: 25%
3	Dissertation progress presentation-2	14th Week from the start date of project semester	Weightage -25%
4	Final project Viva and Dissertation Submission	17th -20th Week of project Semester	Weightage: 30% for Dissertation Weightage: 20% for Final Viva Voce

11. Provision for Appeal

If a candidate is not satisfied with the evaluation of C1, C2 components, he/she can approach the grievance cell with the written submission together with all facts, the assignments, test papers etc, which were evaluated. He/she can do so before the commencement of semester-end examination. The grievance cell is empowered to revise the marks if the case is genuine and is also empowered to levy penalty as prescribed by the university on the candidate if his/her submission is found to be baseless and unduly motivated. This cell may recommend taking disciplinary/corrective action on an evaluator if he/she is found guilty. The decision taken by the grievance cell is final.

For every program there will be one grievance cell. The composition of the grievance cell is as follows:-

- The Controller of Examination - Ex-officio Chairman / Convener
- One Senior Faculty Member (other than those concerned with the evaluation of the course concerned) drawn from the school / department/discipline and/or from the sister schools / departments/sister disciplines – Member.
- One Senior Faculty Members / Subject Experts drawn from outside the University school / department – Member.

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12. Eligibility to Appear Semester End Examination (SEE)

12.1. Only those students who fulfil a minimum of 75% attendance in aggregate of all the courses including practical courses / field visits etc, as part of the course(s), as provided in the succeeding sections, shall be eligible to appear for SEE examination.

12.2. Requirements to Pass a Course

Students are required to score a total minimum of 40% (Continuous Internal assessment and SEE) in each course offered by the University/ Department for a pass (other than online courses) with a minimum of 20 (40% of 50) marks in final examination

13. Requirements to Pass the Semester

To pass the semester, a candidate has to secure minimum of 40% marks in each subject / course of the study prescribed in that semester.

13.1 Provision to Carry Forward the Failed Subjects / Courses:

A student who has failed in a given number of courses in odd and even semesters of first year shall move to third semester of second and final year of the study. However, he / she shall have to clear all courses of all semesters within the double duration, i. e., within four years of admission of the first semester failing which the student has to re-register to the entire program.

13.2 Provision for Supplementary Examination

In case a candidate fails to secure a minimum of 25% (13 marks) in Semester End Examination (SEE) and a minimum of 40% marks together with IA and SEE to declare pass in the course, such candidate shall seek supplementary examination of only such course(s) wherein his / her performance is declared unsuccessful. The supplementary examinations are conducted after the announcement of even semester examination results. The candidate who is unsuccessful in a given course(s) shall appear for supplementary examination of odd and even semester course(s) to seek

for improvement of the performance.

13.3. Provision to Withdraw Course:

A candidate can withdraw any course within ten days from the date of notification of final results. Whenever a candidate withdraws a course, he/she has to register for the same course in case it is hard core course, the same course or an alternate course if it is Soft Core Course or Open Elective Course. A DROPPED course is automatically considered as a course withdrawn.

13.4. Re-Registration and Re-Admission:

a) In case a candidate's class attendance in aggregate of all courses in a semester is less than 75% or as stipulated by the University, such a candidate is considered as dropped the semester and is not allowed to appear for end semester examination (C3) and he / she shall have to seek re-admission to that semester during subsequent semester / year within a stipulated period.

b) In such case where in a candidate drops all the courses in a semester due to personal reasons, it is considered that the candidate has dropped the semester and he / she shall seek re-admission to such dropped semester.

14. Attendance Requirement:

14.1 All students must attend every lecture, tutorial and practical classes.

14.2 In case a student is on approved leave of absence (Example: representing the university in sports, games or athletics, placement activities, NCC, NSS activities and such others) and / or any other such contingencies like medical emergencies, the attendance requirement shall be minimum of 75% of the classes taught.

a) Any student with less than 75% of attendance in aggregate of all the courses including practical courses / field visits etc, during a semester shall not be permitted to appear to the end semester (C4) examination and such student shall seek re-admission as provided in 7.8.4.

b) Teachers offering the courses will place the above details in the School Board meeting during the last week of the semester, before the commencement of C3, and subsequently a notification pertaining to the above will be brought out by the Director of the School before the commencement of C3 examination. A copy of this notification shall also be sent to the office of the Registrar &

Controller of Examination.

15. Absence during Mid Semester Examination:

In case a student has been absent from a mid-semester (C1, C2) examination due to the illness or other contingencies he/she may give a request along with necessary supporting documents and certification from the concerned class teacher/authorized personnel to the concerned Head of the School, for make-up examination. The Head of the School may consider such request depending on the merit of the case and after consultation with course instructor and class teacher, and arrange to conduct a special test for such candidate(s) well in advance before the C3 examination of that respective semester. Under no circumstances C1, C2 test shall be held after C3 examination.

16. Grade Card and Grade Point

16.1. Provisional Grade Card: The tentative/provisional grade card will be issued by the Controller of Examination at the end of every semester indicating the courses completed successfully. The provisional grade card provides **Semester Grade Point Average** (SGPA).

16.2. Final Grade Card: Upon successful completion of M.Sc., Degree a Final Grade card consisting of grades of all courses successfully completed by the candidate will be issued by the Controller of Examination.

16.3. The Grade and the Grade Point: The Grade and the Grade Point earned by the candidate in the subject will be as given below.

Marks P	**Grade** G	**Grade Point** (GP=V x G)	**Letter Grade**
90 -100	10	v*10	O
80 - 89	9	v*9	A+
70 - 79	8	v*8	A
60 - 69	7	v*7	B+
55 - 59	6	v*6	B
50 - 54	5.5	V*5.5	C +
40 - 49	5	v*5	P
0 - 39	0	v*0	F
ABSENT			AB

O - Outstanding; A+-Excellent; A-Very Good; B+-Good; B-Above Average; C+-Average; C-Satisfactory; F – Unsatisfactory.

Here, P is the percentage of marks (P=[C1+C2+C3]) secured by a candidate in a course which is **rounded to nearest integer**. V is the credit value of course. G is the grade and GP is the grade point

16.3.1. Computation of SGPA and CGPA

The Following procedure to compute the Semester Grade Point Average (SGPA)
The SGPA is the ratio of sum of the product of the number of credits with the grade points scored by a student in all the courses taken by a student and the sum of the number of credits of all the courses undergone by a student in a given semester, i.e:

SGPA (Si) = ∑(Ci x Gi) / ∑Ci where Ci is the number of credits of the ith course and Gi is the grade point scored by the student in the ith course.

Illustration for Computation of SGPA and CGPA

Illustration No. 1

Course	Credit	Grade Letter	Grade Point	Credit Point (Credit x Grade)
Course 1	4	A+	9	4X9=36
Course 2	4	A	8	4X8=32
Course 3	3	B+	7	3X7=21
Course 4	3	O	10	3X10=30
Course 5	3	P	5	3X5=15
Course 6	3	B	6	3X6=18
Course 7	2	O	10	2X10=20
Course 8	2	A	8	2X8=16
	24			188

Thus, **SGPA = 188 ÷ 24 = 7.83**

Illustration No. 2

Course	Credit	Grade letter	Grade Point	Credit Point (Credit x Grade point)
Course 1	4	A	8	4X8=32
Course 2	4	B+	7	4X7=28
Course 3	3	A+	9	3X9=27
Course 4	3	B+	7	3X7=21
Course 5	3	B	6	3X6=18
Course 6	3	P	5	3X5=15
Course 7	2	B+	7	2X7=21
Course 8	2	O	10	2X10=20
	24			175

Thus, **SGPA = 175 ÷ 24 = 7.29**

Illustration No.3

Course	Credit	Grade Letter	Grade Point	Credit Point (Credit x Grade point)
Course 1	4	O	10	4 x 10 = 40
Course 2	4	A+	9	4 x 9 = 36
Course 3	3	B+	7	3 x 7 = 21
Course 4	3	B	6	3 x 6 = 18
Course 5	3	A+	9	3 x 9 = 27
Course 6	3	B+	7	3 x 7 = 21
Course 7	2	A+	9	2 x 9 = 18
Course 8	2	A+	9	2 x 9 = 18
	24			199

Thus**, SGPA = 199 ÷ 24 = 8.29**

Cumulative Grade Point Average (CGPA):

Overall Cumulative Grade Point Average (CGPA) of a candidate after successful completion of the required number of credits (96) for Two year Post Graduate degree program is calculated taking into account all the courses undergone by a student over all the semesters of a program i. e.,
CGPA = $\sum(Ci \times Si) / \sum Ci$

Where Si is the SGPA of the ith semester and Ci is the total number of credits in that semester.

The SGPA and CGPA shall be rounded off to 2 decimal points and reported in the transcripts.

Illustration: No.1

CGPA after Final Semester

Semester (ith)	**No. of Credits (Ci)**	**SGPA (Si)**	**Credits x SGPA (Ci X Si)**
1	24	6.83	24 x 6.83 = 163.92
2	26	7.71	26 x 7.71 = 200.46
3	26	8.68	26 x 8.68 = 225.68
4	14	9.20	14 x 9.20 = 128.8
Cumulative	90		718.86

Thus, CGPA = $\frac{24x6.83+26x7.71+26x8.68+14x9.20}{90} = \frac{718.86}{90} = 7.99$

16.3.2. CONVERSION OF GRADES INTO PERCENTAGE:

Conversion formula for the conversion of CGPA into Percentage is:

Percentage of marks scored = CGPA Earned x 10

Illustration: CGPA Earned 8.10 x 10=81.0

16.3.3. Classification of Results

The final grade point (FGP) to be awarded to the student is based on CGPA secured by the candidate and is given as follows.

CGPA	**Grade (Numerical Index)**	**Letter Grade**	**Performance**	**FGP**
	G			**Qualitative Index**
9 >= CGPA 10	10	O	Outstanding	Distinction
8 >= CGPA < 9	9	A+	Excellent	
7 >= CGPA < 8	8	A	Very Good	First Class
6 >= CGPA < 7	7	B+	Good	
5.5> = CGPA < 6	6	B	Above average	Second Class
> 5 CGPA < 5.5	5.5	C	Average	
> 4 CGPA <5	5	P	Pass	Satisfactory

Overall percentage=10*CGPA

17. Challenge Valuation

For all PG courses since it is a double valuation (Internal and External Examiners), candidate shall not have an option to apply for challenge valuation.

Mapping of PEOS with Respect to POs and PSOs

	PO1	P2	PO3	PO4	PO5	PO6	P7	PO8	PO9	PO 10	PSO 1	PSO 2	PSO 3
PEO1	√	√	√	√	√	√	√	√	√	√	√	√	√
PEO2	√	√	√	√	√	√	√	√	√	√	√	√	√
PEO3	√	√	√	√	√	√	√	√	√	√	√	√	√

Mapping of Course Outcomes with programme Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0101	CO1	3	3	2	3			2	1		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	3
	CO3	3	2	2	2			2			3	3	2	3
	CO4	3	3	2	3	2	2	2	1		3	3	2	3
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0102	CO1	3	3	3	3			1	1	1	3	3	3	3
	CO2	3	3	3	2		1	1		1	3	3	2	2
	CO3	3	3	2	2			1		1	3	3	2	2
	CO4	3	3	2	2		2	2	1	1	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0103	CO1	3	3	3	3			1	1		3	3	3	3
	CO2	3	3	2	2	1	1	1			3	3	2	2
	CO3	3	3	2	2	1		1	2	2	3	3	2	2
	CO4	3	3	2	2	1	1	2	1	3	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS111	CO1	3	3	3	3			2	2		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	2
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	2	3	1	1	2	2	2	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS112	CO1	3	3	3	3			2	2		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	2
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	2	3	1	1	2	2	2	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS121	CO1	3	3	3	3			2	2		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	2
	CO3	3	2	2	2			2			3	3	2	2

	CO4	3	3	2	3	1	1	2	2	2	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS122	CO1	3	3	3	3			2	2		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	2
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	2	3			2	2	2	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0104	CO1	3	3	3	3			2			3	3	3	3
	CO2	3	3	3	2			2			3	3	3	3
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	3	3			2			3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0105	CO1	3	3	3	3			2			3	3	3	3
	CO2	3	3	3	2			2			3	3	3	3
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	3	3			2			3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0106	CO1	3	3	3	3			2			3	3	3	3
	CO2	3	3	3	2			2			3	3	3	3
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	3	3			2			3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0201	CO1	3	3	3	2			2		2	3	3	3	2
	CO2	3	3	3	2			2		2	3	3	2	2
	CO3	3	3	2	2			2		2	3	3	2	2
	CO4	3	3	2	2			2		2	3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0202	CO1	3	3	3	3		3	3			3	3	2	2
	CO2	3	3	2	2		2	2			3	3	2	2
	CO3	3	3	2	2		2	2			3	3	2	2
	CO4	3	3	2	2		2	2			3	3	2	2

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0203	CO1	3	3	2	3	2		2		1	3	3	3	2
	CO2	3	3	2	2	3		2		1	3	3	3	2
	CO3	3	3	2	2	3		2		1	3	3	3	2
	CO4	3	3	2	2	3		2		1	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS211	CO1	3	3	3	3		2	3		1	3	3	2	2
	CO2	3	2	2	2		2	3		1	3	3	2	2
	CO3	3	2	2	2			3		1	3	3	2	2
	CO4	3	3	2	2		2	2		1	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS212	CO1	3	3	3	3		2	3		1	3	3	2	2
	CO2	3	2	2	2		2	3		1	3	3	2	2
	CO3	3	2	2	2			3		1	3	3	2	2
	CO4	3	3	2	2		2	2		1	3	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0204	CO1	3	3	2	2	2	1	1	1	1	3	3	2	2
	CO2	3	2	2	2	2	1	1		2	2	3	2	2
	CO3	3	2	2	2	2		1		1	2	3	2	2
	CO4	3	2	2	2	1	1	2	1	1	2	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0205	CO1	3	3	2	2	2	1	1	1	1	3	3	2	2
	CO2	3	2	2	2	2	1	1		2	2	3	2	2
	CO3	3	2	2	2	2		1		1	2	3	2	2
	CO4	3	2	2	2	1	1	2	1	1	2	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0206	CO1	3	3	2	2	2	1	1	1	1	3	3	2	2
	CO2	3	2	2	2	2	1	1		2	2	3	2	2
	CO3	3	2	2	2	2		1		1	2	3	2	2
	CO4	3	2	2	2	1	1	2	1	1	2	3	2	2

M21SL0301	**POS/ COs**	PO1	PO 2	P O 3	PO 4	PO 5	P O 6	PO 7	PO 8	PO 9	PO10	PS0 1	PS0 2	PS0 3
	CO1	2	3		3	2		3				3	3	2
	CO2	3	2									2	2	3
	CO3	3	2		3			3		3	3	3	3	2
	CO4	2	3	3	3			3				3	3	3
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z01	CO1	3	3	2	3	1	3	1	2	1	3	3	2	2
	CO2	3	2	2	3	1	3	1	2	1	2	3	2	2
	CO3	3	2	2	3		2	1	2	2	2	3	2	2
	CO4	3	3	2	3	1	2	2	2	1	2	3	2	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z02	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z03	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z04	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z05	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z06	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z07	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z08	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z09	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z10	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z11	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z12	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0Z13	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z14	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z15	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z16	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z17	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z18	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS311	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS312	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SLO301	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SLO302	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0401	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOON01	CO1	2	2	3			3					1	1	3
	CO2	3	2									1	1	3
	CO3	3	3								3	1	1	3
	CO4	3	3								3	1	1	3
Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOON02	CO1	2	2	3			3					1	1	3
	CO2	3	2									1	1	3
	CO3	3	3								3	1	1	3
	CO4	3	3								3	1	1	3
Course Code	POS / COS	PO1	PO 2	P O 3	PO 4	PO 5	P O 6	PO 7	PO 8	PO 9	PO10	PS O1	PS O2	PS O3
M21SO0401	CO1	2	2	3			3					1	1	3
	CO2	3	2									1	1	3
	CO3	3	3								3	1	1	3
	CO4	3	3								3	1	1	3

REVA UNIVERSITY

School of Applied Sciences

Department of Biotechnology

MSc (Microbial Technology) Program

Scheme of Instruction (Effective from Academic Year 2021-22)

FIRST SEMESTER	**Course code**	**Course**	**HC/ SC/ OE**	**L**	**T**	**P**	**Credits**	**Hours/ Week**
1	**M21SO0101**	Microbiological methods	HC	4	0	0	4	4
2	**M21SO0102**	Molecular Microbial Genetics	HC	4	0	0	4	4
3	**M21SO0103**	Bioanalytical Techniques	HC	4	0	0	4	4
4	**M21SOS111**	Research Methodology	SC	4	0	0	4	4
5	**M21SOS112**	Entrepreneurship in Life Sciences						
6	**M21SOS121**	Cell Culture Technology	SC	4	0	0	4	4
7	**M21SOS122**	Microbial Metabolites						
	PRACTICAL SESSIONS							
8	**M21SO0104**	Microbiological Methods LAB	HC	0	0	2	2	4
9	**M21SO0105**	Molecular Microbial Genetics LAB	HC	0	0	2	2	4
10	**M21SO0106**	Bioanalytical techniques Lab	HC	0	0	2	2	4
	Total Credits			**20**	**0**	**6**	**26**	**32**

SECOND SEMESTER			HC/ SC/ OE	L	T	P	Credits	Hours/ Week
1	**M21SO0201**	Microbial Metabolism and Physiology	HC	4	0	0	4	4
2	**M21SO0202**	Microbial Diversity	HC	4	0	0	4	4
3	**M21SO0203**	Bioinformatics and Biostatistics	HC	4	0	0	4	4
4	**M21SOS211**	Microbiology for Sustainable Environment	SC	4	0	0	4	4
5	**M21SOS212**	Microbial Food Processing						
	PRACTICAL SESSIONS							
6	**M21SO0204**	Microbial metabolism and Physiology Lab	HC	0	0	2	2	4
7	**M21SO0205**	Microbial Diversity LAB	HC	0	0	2	2	4
8	**M21SO0206**	Bioinformatics and Biostatistics Lab	HC	0	0	2	2	4
9	**M21PTM201**	Soft Skill Training (Mandatory Course) offered by Training and Placement	MC	3	0	0	0	3
	Total Credits			**18**	**0**	**6**	**22**	**28**

THIRD SEMESTER			HC/ SC/ OE	L	T	P	Credits	Hours/ Week
	Specialization in Industrial & Food Microbiology							
1	**M21SOZ301**	Food Microbiology	HC	4	0	0	4	4
2	**M21SOZ302**	Bioprocess Technology	HC	4	0	0	4	4
3	**M21SOZ303**	Food Chemistry and Processing	HC	4	0	0	4	4

4	**M21SOS311**	Nano biotechnology and Space Microbiology	SC	4	0	0	4	4
5	**M21SOS312**	Marine Microbiology						
	PRACTICAL SESSIONS							
6	**M21SOZ304**	Food Microbiology Lab	HC	0	0	2	2	4
7	**M21SOZ305**	Bioprocess Technology Lab	HC	0	0	2	2	4
8	**M21SOZ306**	Food Chemistry and Processing lab	HC	0	0	2	2	4
9	**M21SLO301**	Organic Farming	OE	4	0	0	4	4
10	**M21SL0302**	Skill Enhancement Course	HC	2	0	0	2	2
	Total Credits			**22**	**0**	**6**	**28**	**34**
	Specialization in Agriculture & Environmental Microbiology							
1	**M21SOZ307**	Soil Microbiology	HC	4	0	0	4	4
2	**M21SOZ308**	Plant Pathology	HC	4	0	0	4	4
3	**M21SOZ309**	Biomass and energy systems	HC	4	0	0	4	4
4	**M21SOS311**	Nano biotechnology and Space Microbiology	SC	4	0	0	4	4
5	**M21SOS312**	Marine Microbiology						
	PRACTICAL SESSIONS							
6	**M21SOZ310**	Soil Microbiology Lab	HC	0	0	2	2	4
7	**M21SOZ311**	Plant Pathology Lab	HC	0	0	2	2	4
8	**M21SOZ312**	Biomass and energy systems Lab	HC	0	0	2	2	4
9	**M21SLO301**	Organic Farming	OE	4	0	0	4	2
10	**M21SL0302**	Skill Enhancement Course	HC	2	0	0	2	2
	Total Credits			**22**	**0**	**6**	**28**	**30**
	Specialization in Medical and Pharmaceutical Microbiology							
1	**M21SOZ313**	Clinical Microbiology	HC	4	0	0	4	4
2	**M21SOZ314**	Medical Biochemistry	HC	4	0	0	4	4

3	**M21SOZ315**	Pharmaceutical Microbiology	HC	4	0	0	4	4
4	**M21SOS311**	Nanobiotechnology and Space Microbiology	SC	4	0	0	4	4
5	**M21SOS312**	Marine Microbiology						
	PRACTICAL SESSIONS							
6	**M21SOZ316**	Clinical Microbiology Lab	HC	0	0	2	2	4
7	**M21SOZ317**	Medical Biochemistry Lab	HC	0	0	2	2	4
8	**M21SOZ318**	Pharmaceutical Microbiology Lab	HC	0	0	2	2	4
9	**M21SLO301**	Organic Farming	OE	4	0	0	4	2
10	**M21SL0302**	Skill Enhancement Course	HC	2	0	0	2	2
	Total Credits			**22**	**0**	**6**	**28**	**30**
FOURTH SEMESTER			**HC/ SC/ OE**	**L**	**T**	**P**	**Credits**	**Hours/ Week**
1	**M21SO0401**	Major Project	HC	0	0	10	10	10
2	**M21SOON01**	Swayam course-1	-	-	-	-	2	-
3	**M21SOON02**	Swayam course-2	-	-	-	-	2	-
	Total Credits					**10**	**14**	**10**

Note: Open elective offered by Department of Biotechnology, Organic Farming shall be opted by other schools. MSc Microbial Technology shall have to opt open elective courses offered by other schools.

Semester-wise Summary of Credit Distribution

Semesters	L	T	P	No. of Credits
First Sem	**20**	**0**	**6**	**26**
Second Sem	**18**	**0**	**6**	**22**
Third Sem	**22**	**0**	**6**	**28**
Fourth Sem	**0**	**0**	**10**	**14**
Total Credits	**06**	**0**	**28**	**90**

FIRST SEMESTER

Course code	Microbiological Methods	Course type	L	T	P	C	CH
M21SO0101		HC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have basic knowledge of culturing of various microorganisms.

Course Objectives:

1. Enable students to understand the basic biomolecules and their role in life.
2. Acquire the knowledge of basic and biological functions of immune system.
3. Enhance the analytical skills related to microbiology.
4. To equip students with methodologies to detect and identify the microbes.

Course Outcomes:

By the end of the course the student will be able to:

1. Characterize the basis and importance's of biochemical reactions involved in cells.
2. Apply the basic knowledge and importance's of immune system in organisms.
3. Apply and analyze the techniques to characterize and identify of microbes.
4. Employ the methodologies to culture and analyses the various microbes.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO 1	PO2	PO 3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PSO1	PSO 2	PS O3
M21SO0101	CO1	3	3	2	3			2	1		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	3
	CO3	3	2	2	2			2			3	3	2	3
	CO4	3	3	2	3	2	2	2	1		3	3	2	3

Course Content:

Unit-I Basic concepts of Chemistry and Biochemistry **13 h**

Chemical formulas and the mole concept. Avogadro's constant. Chemical reaction and equations. Mass relationship in reactions & calculations, Titrations and indicators. Scope and importance of biochemistry, Structure of water, Acid base concept and buffers, pH, Hydrogen bonding,

Hydrophobic, Electrostatic and Vander Waals forces. Classification, structure and function of carbohydrates, proteins and lipids. Structure and function of amino acids and vitamins.

Unit-II Microbial life process **13 h**

Bacterial growth and fungal growth- mechanism of reproductions, Measurement of microbial growth, Growth curve and factors influencing the microbial growth. Growth stoichiometry; Kinetics of microbial growth and product formation in batch, Fed batch and continuous cultures. Industrially important microbes and strain improvement.

Unit-III Microscopy and microbiological staining **13 h**

Light microscopy- Simple microscope, Compound microscope (Bright field, Dark field, Phase contrast, Fluorescence and confocal). Electron Microscopy- Principles, construction and mode of operation of Scanning, transmission electron microscope, AFM and STEM, limitations and preparation of specimens. Types of stains and principles of staining in microbiology, wet mounting and hanging drop method.

Unit-IV Culture methods and Aseptic Techniques **13 h**

Microbiological media-types, cultivation of aerobic and anaerobic bacteria, pure cultures and cultural characteristics. Serial dilution, Pure cultures and isolation techniques, Maintenance and preservation of pure cultures. Principles and methods of sterilization-Physical and Chemical Sterilization.

Reference

1. Microbiology; Lansing M Prescott, John P. Harley, Donald A Klein, Sixth edition, Mc Graw Hill Higher education (2005).
2. General Microbiology; RY Ingraham, JL Wheels, M.L. Painter. Thess Macmillan Press Ltd (1992).
3. Brock Biology of Microorganism; M.T, Martinko, J.M. Parker, Prentice-Hall (2006).
4. Microbiology; M.J. Pelczar, E.C.S Chan and N.R. Kreig, Tata MacGraw Hill (2004).
5. Microbial Genetics, S.R. Molloy, J.E. Jr. Cronan and Frreifelder D Jones, Bartiett Publishers.
6. Bergey's Manual of Systematic Bacteriology, Breed and Buchanan, 2nd Edition, (Volumes. 1 – 6) (2001 – 2003).
6. General Microbiology, R. Y. Stanier, E. A. Adelberg, J. L. Ingraham, 4th edition, Mac Millan Press, London (2000).
7. Microbiology An introduction; Tortora Funke case (2001).

Course code	Molecular Microbial Genetics	Course type	L	T	P	C	CH
M21SO0102		HC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students to know basics and importance of DNA and RNA.

Course Objectives:

1. Understand the basics of genetic material and hereditary.
2. Study the mechanism of transcription and translation of genetic information.
3. Understand the mechanism of recombinant and genetic transfer in bacteria
4. Discuss the genome structure and functions bacteriophages and eukaryotic microorganisms.

Course Outcome:

By the end of the course the student will be able to:

1. Outline the genetic inheritance in prokaryotic and eukaryotic microorganisms.
2. Explore the mechanism of transcription and translation in microorganisms.
3. Acquire the knowledge of genetic transfer mechanisms in microorganisms.
4. Exploit the knowledge of microbial genetics in allied research.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PSO1	PSO 2	PSO 3
M21SO0102	CO1	3	3	3	3			1	1	1	3	3	3	3
	CO2	3	3	3	2		1	1		1	3	3	2	2
	CO3	3	3	2	2			1		1	3	3	2	2
	CO4	3	3	2	2		2	2	1	1	3	3	2	2

Course Content:

Unit-I Molecular Basis and life process **13 h**

Nucleic acids as hereditary material and concept of central dogma. Structure and functions of DNA and RNA, Forms of DNA and Types of RNA- rRNA, tRNA and mRNA. RNA Biology-iRNA & miRNA. DNA replication and DNA polymerases of *E. coli* and eukaryotes. Models and enzymes in DNA replication in prokaryotes and eukaryotes. Cell cycle and regulation of DNA replications, molecular mechanism of mutation and DNA repair.

Unit-II Gene Expression and regulation **13 h**

Transcription, RNA polymerases, Types of promoters, Reverse transcriptase and RNA replicase, Genetic code and Translation in prokaryotic and eukaryotic microbes. Post-transcriptional modification, maturation and splicing of RNA transcripts and catalytic RNA. Post-translational modification and protein targeting. Gene regulation at transcriptional & translational level, operon-positive, negative regulation and attenuation (Lac & Trp operon).

Unit-III Gene transfer methods in microorganisms **13 h**

Methods of gene transfer in bacteria-Conjugation-Discovery, nature of donor strains and compatibility, interrupted mating and temporal mapping and molecular pathway of recombination. Transformation-natural transformation systems, Biology of transformation, transformation and gene mapping, Transduction-discovery, generalized and specialized or restricted transduction.

Unit-IV Genetics of phage and fungi **13 h**

T4 virulent phage-structure, life cycle, genetic map and DNA replication. Lambda phage: Structure, genetic map, lytic and lysogenic cycle. Applications of phages in microbial genetics. Transposons-Insertion sequences and composite transposons, phages as transposons, replicative, non-replicative and conservative transposition. Aspects of fungal Genetics: Meiotic and mitotic mapping, gene conversion, mitotic segregation and recombination, heterothallism and mating type switches.

References

1. Genes V; Benjamin Lewin, Oxford University Press, New York (1998).
2. Gene IX; Benjamin Lewin Oxford University Press, New York (2005).
3. Principles of Genetics; Snustad and Simmons, Fourth Edition, John Wiley and Sons, Inc (2006).
4. Molecular Cell Biology; Lodish et.al., W. H. Freeman and Company (2006).
5. Genomes; T.A. Brown, John Wiley and sons (Asia)PTE LTD, New York (2001).
6. Principles of Gene Manipulation and Genomics; S.B. Primrose and R. M. Twyman, Seventh edition, Blackwell Publishing, U.K (2002).
7. Modern Microbial Genetics. 1991; Streips and Yasbin. Niley Ltd (2000)
8. Molecular Biology of the Gene 4th Edition by J.D. Watson, N.H. Hoppkins, J.W (2009)
9. Microbial Genetics by Maloy ET. Al. 1994; Jones and Bartlett Publishers (2000).

Course code	Bioanalytical techniques	Course type	L	T	P	C	CH
M21SO0103		HC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Student should have a knowledge of biomolecules and separation techniques.

Course Objectives:

1. To develop the skills and competency in biochemical techniques
2. To equip with separation of biomolecules through various chromatographic techniques

3. To provide new skills for the separation and characterisation of biomolecules
4. Empower the knowledge of physico-chemistry to analyse the biomolecules

Course Outcome:

By the end of the course the student will be able to:

1. Acquire the basic skill of separation of macro- and micromolecules from biological sources.
2. Develop the biochemical techniques to analyses and quantify the biomolecules.
3. Upgrade the knowledge of biochemical techniques to characterize the biomolecules.
4. Quantitative analysis of biomolecules by various physicochemical methods.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0103	CO1	3	3	3	3			1	1		3	3	3	3
	CO2	3	3	2	2	1	1	1			3	3	2	2
	CO3	3	3	2	2	1		1	2	2	3	3	2	2
	CO4	3	3	2	2	1	1	2	1	3	3	3	2	2

Course Content:

Unit -1 Centrifugation **13 h**

Principles of separation techniques, Centrifugation-classification, types and applications, High speed and Ultracentrifugation, Differential and Density-gradient centrifugation, Analytical centrifugation and applications, Factors affecting Sedimentation, Preparative and analytical centrifugation, Safety measures of centrifugation.

Unit -II Chromatography **13 h**

Chromatography techniques classification, factors affecting the separation, Thin layer chromatography, Column chromatography, gel permeation chromatography, ion exchange chromatography, Affinity chromatography, HPLC, GC, applications

Unit -III Electrophoresis **13 h**

Agarose gel electrophoresis, factors affecting separation, troubleshooting, Poly Acrylamide gel electrophoresis, SDS-PAGE, 2D-PAGE; capillary electrophoresis; Isoelectric focusing and isotachophoresis, Hybridization and blotting techniques.

Unit -IV Spectroscopy and Radioactivity **13 h**

Electromagnetic spectrum and interaction of radiation with matter, UV and Visible spectroscopy; Beer-Lambert's law and its limitation; IR spectroscopy- Principles, NMR and ESR, CD/ORD, X-ray crystallography.Mass spectrometry- MALDITOF, TANDEM Mass spectrometry, GC-MS and

LC- MS. Radioactivity - Introduction to Isotopes, detection and measurement of radioactivity, Autoradiography, Radio-labeling Techniques and applications.

References

1. Microbiology; M.J. Pelczar, E.C.S Chan and N.R. Kreig, Tata MacGraw Hill (2004).
2. Biochemical techniques; Manikam and Sadasivam, New Age International, 2008.

Course code	Research Methodology	Course type	L	T	P	C	CH
M21SOS111		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should be knowing about the research ideas and recent ongoing research works.

Course Objectives:

1. Acquire basic knowledge on qualitative and quantitative methods.
2. Identify and formulate the research problems and design the research.
3. Familiarize the student in using statistical tools in research methods.
4. Student will acquire the knowledge biological information and relevant ethical concerns.

Course Outcome:

By the end of the course the student will be able to:

1. Understand the fundamentals of research and research writings
2. Identify the statement of problem, research gaps and study design in the research
3. Analyze the parametric and non-parametric research data types, sampling methods and use of statistical tools in study design.
4. Apply the data collections, databases and regulations in conducting research.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0111	CO1	3	3	3	3			2	2		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	2
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	2	3	1	1	2	2	2	3	3	2	2

Course Content:

Unit I Fundamental concepts in Research **13 h**

Concept of theory, empiricism, deductive and inductive theory, types of research-qualitative and quantitive research. Utility for research, characteristics of scientific methods-understanding the language of research and language in scientific research.

Unit II Research Problem and Research design **13 h**

Scientific Research-problem, definition, objectives, types, purposes and components of research problem. concept and importance in research-features of a good research design, exploratory research design and descriptive research designs.

Unit III Sampling, data analysis and reporting in research **13 h**

Sampling methods and sampling errors, determining size of the sample. Experimental design-concept of independent & dependent variables. Fundamentals of statistical analysis, correlation and regression. Research reports-structure, components, types and layout of research report and articles, writing and interpreting research results, figures and graphs.

Unit IV Tools in research and Regulatory considerations **13 h**

Research Guides, Handbook, Academic Databases for Biological Science Discipline. Methods to search required information effectively. Data mining tools, searching and bioinformatics databases. Guidelines-animal ethical committee, animal models, various routes of drug administrations, LD50, ED50 and EPA for environment.

References

1. Introduction to research methods: a practical guide for anyone undertaking a research project, Oxford: How To Books; Catherine Dawson, Reprint (2010).
2. Random Data: Analysis and Measurement Procedures, 4th Edition; Julius S. Bendat, Allan G. Piersol (2011)
3. Research in Medical and Biological Sciences, 1st Edition, From Planning and Preparation to Grant Application and Publication, Editors: Petter Laake Haakon Benestad Bjorn Olsen, (2015).
4. Research Design: Qualitative, Quantitative, and Mixed Methods Approaches, Fourth Edition; John Creswell (2013).

Course code	Entrepreneurship in Life Sciences	Course type	L	T	P	C	CH
M21SOS112		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should be knowing about business in life sciences and important facts.

Course Objectives:

1. Acquire the knowledge of entrepreneur skills related to life science.
2. Opportunities of business opportunities in various fields of life science.

3. Inculcate the ideas of business development and commercialisation of products/services.
4. Familiarise about the regulations and protection of intellectual property right related to business.

Course Outcome:

By the end of the course the student will be able to:

1. Differentiate the various entrepreneurial skills related to life science.
2. Classify the parameter to access opportunities and constrains, ideas in startups.
3. Explore the systematic process of select and screen of novel business ideas
4. Understand the regulatory bodies in protecting the business.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PSO1	PSO 2	PSO 3
M21SOS112	CO1	3	3	3	3			2	2		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	2
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	2	3	1	1	2	2	2	3	3	2	2

Course Content:

Unit I Fundamentals of Bio-business **13 h**

Concept of Bio-business, contemporary vs antique, bio-business, wealth creation in bio-business, healthcare, biomedical sciences, industrial life sciences and biotechnology, quick survey of regional and global strengths and capabilities, Startups related opportunities, company registration and funding opportunities.

Unit II Life Science based bio-business **13 h**

Agriculture based business, food industry, business related to environment management, bioremediation, bioleaching and waste management. Case studies of business failure and success stories, biotech clusters, process of business commencement, ethics in business, reason for business failures, causes and preventive measures.

Unit III Entrepreneurship development **13 h**

Myths of Entrepreneurship, factors affecting entrepreneurship growth, future of entrepreneurship, entrepreneurship development programs (EDP) technology business incubator, open discussion recent updates, group project presentation: case studies of different industries and their strategic planning.

Unit IV Protection of IP in bio-business **13 h**

Opportunities for business, Policy and Regulatory Concerns, Human Resource, Financing incentives and subsidies & bounties for business units by government and NGOs. Intellectual Property, Technology Licensing and Branding, Patenting-country of origin and international origin, Copyright, Geographical Indicator, Trade Secret and case studies.

Reference

1. 101 Real Life Business Lessons for Emerging Entrepreneurs Hardcover; Bill Green, Publisher: Koehler Books (2017).
2. The Tactics, Routines, and Habits of Billionaires, Icons, and World-Class Performers Hardcover; Timothy Ferriss, Arnold Schwarzenegger., (2016).
3. My Misadventure in the Start-Up Bubble Hardcover; Dan Lyons, Publisher: Hachette Books (2016).
4. An Entrepreneur's Vision of the Future Hardcover, Steve Case Publisher (2016).
5. The Only Move That Matters Is Your Next One Hardcover, Jenny Blak (2016).

Course code	Cell Culture Technology	Course type	L	T	P	C	CH
M21SOS121		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should be knowing about the culturing of cells and its importance in research.

Course Objectives:

1. Explain the different types of cell lines in animal cell culture techniques
2. Deep understanding of the composition and preparation methodology of the different types of cell culture media.
3. Illustrate the importance and methodology for transferring genes into the different types of animal and plant cells.
4. Apply the knowledge for a better understanding of the different explant-based plant tissue culture callus formation and as suspension culture methods.

Course Outcome:

By the end of the course the student will be able to:

1. Elaborate the fundamentals of animal cell culture and types of media used in animal cell culture.
2. Classify the different types of stem cells as well as compare and contrast the various types of stem cells and outline the different cord blood-based stem cell techniques.
3. Extend and apply the fundamental knowledge for the production of animal and plant tissue culture- based products.
4. Outline the methods involved in gene transfer techniques in animal and plant cells.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PO9	PO10	PSO1	PSO 2	PSO 3
M21SOS121	CO1	3	3	3	3			2	2		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	2
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	2	3	1	1	2	2	2	3	3	2	2

Course Content:

Unit I Cell culture techniques **13 h**

Essentials of animal cell culture-Animal cell culture, primary cells, cells lines. Adherent and non-adherent cells. Resources of cell lines and viability testing. Media and methods to culture animal cells. Subculturing and maintenance of animal cells. Medium preparation and methods to avoid contamination. Continuous and batch mode of large-scale cell culture. 2D and 3D culture methods. Methods for encapsulation of animal cells and its advantages. Methods for Immortalization of primary cells and industrial applications of cell culture.

Unit II Stem cell technology **13 h**

Embryonic stem cells, cord blood stem cells, Induced pluripotent stem cells. Difference between Totipotent, Multipotent and pluripotent stem cells. Methods for obtaining the stem cells and their advantages with drawbacks. Methods for tissue development using in vitro cultured cells.

Unit III Plant tissue and cell culture **13 h**

Different explants used for plant tissue culture with advantages and disadvantages. Callus and suspension culture. Methods to induce callus from explants, somatic embryos. MS medium, shoot medium and root medium. Methods in plant cell culture for crop improvement. Production of metabolites using plant cell suspension culture, Bioelicitation studies, Industrial application of Plant cell culture.

Unit IV Gene transfer techniques in to host cells **13 h**

In Animal cells- Transfection, Transduction, electroporation. Transient and stable expression of transgene in animal cells and Plant Cells- Protoplast fusion technique and its advantages. Methods for gene transfer in plant cells and generation of transgenic plants. Selection and identification techniques of transformants in plants and animal cells.

References

1. Culture of Animal Cells: A Manual of Basic Technique and Specialized Applications, Sixth Edition; R. Ian Freshney, Elsevier, (2011).
2. Plant Cell Culture: Essential Methods; Michael R. Davey, Paul Anthony, Springer (2010).

3. Plant Cell Culture Protocols, Loyola-Vargas, Víctor M., Ochoa-Alejo, Springer (2012).
4. Embryonic Stem Cells; John R Masters, Bernhard O Palsson, James A Thomson. 6th Volume, Springer (2010).

Course code	Microbial Metabolites	Course type	L	T	P	C	CH
M21SOS122		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have basic knowledge of commercial products from microbes.

Course Objective:

1. Acquire the knowledge of various metabolites from microbes and their applications
2. Describe and design a microbial production of primary metabolites from microbes.
3. Inculcate the various microbial metabolites and their importance
4. Familiarise about the value-added products from the microbes.

Course Outcome:

By the end of the course the student will be able to:

1. Productively translate both basic and frontiers research concepts relating to metabolite production and purification into a modern industrial bioprocess perspective.
2. Describe and analyse the control of in vitro cellular growth processes within the industrial scale bioreactor environment,
3. Demonstrate a technical lexicon that will allow productive interface with complementary disciplines.
4. Discuss and evaluate the operational considerations and downstream processing of biotechnology products.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SOS122	CO1	3	3	3	3			2	2		3	3	3	3
	CO2	3	2	2	2			2			3	3	2	2
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	2	3	1	1	2	2	2	3	3	2	2

Course Content:

Unit I Microbial products as primary and secondary metabolites 13 h

Metabolites- trophophase- Ideophase relationships in production of secondary metabolite; Role of secondary metabolites in physiology of organisms producing them; Pathways for the synthesis of primary and secondary metabolites of commercial importance; Metabolic control mechanisms: substrate induction; catabolic regulation; feedback regulation; amino acid regulation of RNA synthesis; Energy charge regulation and permeability control; Bypassing/ disorganization of regulatory mechanisms for overproduction of primary and secondary metabolites

Unit II Microbial Primary metabolites production 13 h

Fermentation process and phases of metabolite production, metabolic pathways and their fermentation products. Organic feedstock: ethanol; Acetone; Ethanol Organic acids: Production of Citric acid; Acetic acid; Lactic acid; Gluconic acid; Kojic acid; itaconic acid; Amino acids: Use of amino acids in industry; methods of production; Production of individual aminoacids (L-Glutamic acid; L Lysin; L-Tryptophan).

Unit III Microbial enzymes 13 h

Enzymes-classification, nomenclature and commercial applications of enzymes in various fields, production of Amylases; Glucose Isomerase; L Asparaginase Proteases Renin; Penicillin acylases; Lactases; Pectinases; Lipases; Structure and biosynthesis Nucleosides Nucleotides and related compounds.

Unit IV Microbial Vitamins and value-added products 13 h

Vitamins-Vitamin B12; Riboflavin; B carotene; Antibiotics: beta-Lactam antibiotics; amino acid and peptide antibiotics; Carbohydrate antibiotics; Tetracycline and antracyclines; Nucleoside antibiotics; Aromatic antibiotics; bioplastics (PHB; PHA); biotransformation of steroids.

References

1. A Textbook of Industrial Microbiology; W. Crueger and A. Crueger. Publisher: Sinauer Associates (2010).
2. Industrial microbiology, G. Reed, Publishers: CBS (2010).
3. Biology of Industrial microorganisms; A. L. Demain (2012).
4. Principles of fermentation technology; Stanbury P.F.A. Whitaker and Hall (2012).
5. Fermentation and Biochemical Engineering Handbook: Principles, Process Design, and Equipment; H.C. Vogel, C.L. Todaro, C.C. Todaro. Publisher: Noyes Data Corporation/ Noyes Publications (2015).
6. New Products and New Areas of Bioprocess Engineering (Advances in Biochemical Engineering/Biotechnology, 68) T. Scheper. Publisher: Springer Verlag (2012).

Course code	Microbiological methods LAB	Course type	L	T	P	C	CH
M21SO0104		HC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should be aware of various culture media and culturing aspects of microorganisms.

Course Objectives:

1. Practical exposure in isolation of microorganisms from different samples
2. Analyse various staining and quantification procedures used in microbiology
3. Determine the biochemical analysis of carbohydrates, proteins and fats
4. Estimation and preparations of the reagents and solutions for biochemical assay

Course Outcomes:

By the end of the course the student will be able to:

1. Hands on exposures to isolate and quantify the microorganisms
2. Exploit the staining protocols used in the microbiology
3. Quantify the carbohydrates and protein in the various samples
4. Exploit the preparation and analysis of reagents and buffers for biochemical analysis.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO0104	CO1	3	3	3	3	2		2		2	3	3	3	3
	CO2	3	3	3	2	2		2		2	3	3	3	3
	CO3	3	2	2	2	2		2		2	3	3	2	2
	CO4	3	3	3	3	2		2		2	3	3	2	2

Course Content:

1. Isolation of microflora from air, soil and water.
2. Simple staining techniques in microbiology.
3. Differential staining techniques in microbiology.
4. Sugar fermentation tests, urease test, triple sugar iron test, oxidase, lipid, DNAse, catalase and casein hydrolysis test.
5. Enumerating the yeast cells by the hemocytometer

6. Preparation of standard and buffer solutions.
7. Estimation of microbial total sugars by anthrone method.
8. Estimation of microbial reducing sugars by DNS method.
9. Estimation of Microbial proteins by Lowrys' method
10. Estimation of saponification and iodine value of oils and fats from microbes

Reference

1. Analytical techniques in Biochemistry and molecular Biology; Katoch, Rajan Springer (2011)
2. Basic Methods for the Biochemical Lab; Martin Holtzhaue, Springer (2007).
3. Principles and Techniques of Biochemistry and Molecular Biology; Keith Wilson and John Walker, 7th edition Cambridge University Press (2010).
4. Experiments in Applied Microbiology Samuel Singer; Academic Press (2001).
5. Microbiological methods; Collins, C.H., Tatrica M. Lyne and Grange, J.M. 8th edition. Hodder Arnold publishers (2004).

Course code	Molecular Microbial Genetics LAB	Course type	L	T	P	C	CH
M21SO0105		HC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have basic knowledge of isolation of DNA from cells.

Course Objectives:

1. Practical knowledge on the DNA and RNA analysis
2. Determine the DNA and RNA in the various samples
3. Understand the bacterial genetic process occurring in nature
4. Inculcate the molecular mechanism for the research uses.

Course Outcomes:

By the end of the course the student will be able to:

1. Practical approaches in the DNA and RNA analysis
2. Estimate the DNA and RNA in the various samples
3. Demonstrate the bacterial process happening in nature
4. Experiment the molecular analysis of research purpose

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO0105	CO1	3	3	3	3			2			3	3	3	3
	CO2	3	3	3	2			2			3	3	3	3

	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	3	3			2			3	3	2	2

Course Contents:

1. Isolation of bacterial genomic DNA and analysis by AGE
2. Isolation of plasmid DNA and analysis by AGE
3. Estimation of DNA by DPA method.
4. Estimation of RNA by orcinol method.
5. Bacterial transformation by α-complementation.
6. Demonstration of Bacterial conjugation
7. Demonstration of Bacterial transduction
8. Isolation of streptomycin resistant strain of *E. coli* by gradient plate method
9. Replica plating technique for transfer of bacterial colonies.
10. Induction of mutation in the *E coli* by EMS

Reference

1. Molecular Biology of the Cell; Bruce Alberts, Alexander Johnson, Julian Lewis, David Morgan, Martin Raff, Keith Roberts, Peter Walter. 6th Edition., Garland Science publisher (2014).
2. Molecular Cell Biology; Harvey Lodish, Arnold Berk, Chris A Kaiser, Monty Krieger, Anthony Bretscher, Hidde Ploegh, Angelika Amon, Matthew P Scott. 7th edition Macmillan Publishers (2012).
3. The Cell: A molecular approach; Geoffrey M. Cooper and Robert E. Hausman. 7th edition. Publisher: Sinauer Associates, USA (2016).

Course code	**Bioanalytical techniques LAB**	**Course type**	**L**	**T**	**P**	**C**	**CH**
M21SO0106		**HC**	**0**	**0**	**4**	**2**	**4**

Prerequisites/Pre reading for the course:

Students should have theoretical knowledge of separation and analysis of biomolecules.

Course Objectives:

1. Hands on experience in the preparation of reagents and buffers
2. Acquire analytical skills in determination of molecular weight of microbial proteins
3. Exploit the various advanced skills for separation and characterisation of biomolecules.
4. Optimization of protein purification through various methods.

Course Outcomes:

By the end of the course the student will be able to:

1. Equipped with biochemical techniques for handling the biomolecules.
2. Apply the knowledge of practical skills in separation of biomolecules
3. Explore various methods to characterize the biomolecules.
4. Quantify the biomolecules with optimized methods.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0106	CO1	3	3	3	3			2			3	3	3	3
	CO2	3	3	3	2			2			3	3	3	3
	CO3	3	2	2	2			2			3	3	2	2
	CO4	3	3	3	3			2			3	3	2	2

Course Contents:

1. pH metric titrations for determining p Ka of a weak acid for optimizing buffer capacity
2. Separation of bacterial proteins by paper chromatography
3. Separation of bacterial proteins by TLC
4. Separation of extracts by Silica gel column chromatography.
5. Separation of proteins by sepharose column chromatography.
6. Separation of Immunoglobulin by affinity chromatography.
7. Molecular weight detection of separated proteins by silver staining of SDS PAGE
8. Molecular weight detection of separated proteins CBB staining by SDS PAGE.
9. Demonstration of separation of metabolites by HPLC
10. Demonstration of analysis of GC-MS.

References

1. Principles of Biochemistry, Lehninger and D L Nelson, 6th edition. Macmillan Publications (2012)
2. Genes XI, Lewin, Benjamin; Krebs, Jocelyn E.; Goldstein, Elliott S.; Kilpatrick, Stephen T., Jones and Bartlett Learning (2014).
3. Biochemistry, Jeremy M Berg, John L Toymoczko and LubertStryer , 8th Edition Macmillan Publications (2015).

SECOND SEMESTER

Course code	Microbial Metabolism and Physiology	Course type	L	T	P	C	CH
M21SO0201		HC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should be aware of basic concepts of microbial process in the nature.

Course Objectives:

1. Gain the knowledge of microbial photosynthesis and their extreme adaptations.
2. Understand the microbial metabolic pathway occurring in the nature.
3. Explore the better understanding of lipid and nucleic acid metabolism.
4. Exploit the biological nitrogen fixation for other applications.

Course Outcomes:

By the end of the course the student will be able to:

1. Explain the various pathways involved in microbial photosynthesis
2. Utilize the microbial metabolic pathways for commercial uses
3. Exploit the various metabolic process in degradation of lipids and nucleic acids
4. Explore various nitrogen fixation mechanism for commercial uses

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO0201	CO1	3	3	3	3		3	3			3	3	2	2
	CO2	3	3	2	2		2	2			3	3	2	2
	CO3	3	3	2	2		2	2			3	3	2	2
	CO4	3	3	2	2		2	2			3	3	2	2

Course Contents:

Unit-I Microbial photosynthesis **13 h**

Brief account of microbial photosynthesis – oxygenic and anoxygenic photosynthesis; fixation of CO_2-Calvin cycle-C3-C4 pathway. Chemolithotrophy – sulphur, iron, hydrogen, nitrogen oxidations, methanogenesis and luminescence.

Unit-II Microbial metabolism of carbohydrates **13 h**

Respiratory metabolism-glycolysis, Entner Doudroff pathway, glyoxalate pathway, Krebs cycle-oxidative and substrate level phosphorylation-reverse TCA cycle. Fermentation-homo and heterolactic fermentation. Enzymes-classification, mechanism of action; Factors affecting enzyme action and Immobilized enzymes.

Unit-III Metabolism of lipids and nucleic acids **13 h**

Lipid metabolism: β-oxidation, Biosynthesis of fatty acids, degradation of fatty acids. Synthesis of purines and pyrimidines-de novo and salvage pathways. Biosynthesis of glycoproteins and peptidoglycan.

Unit-IV Nitrogen Metabolism **13 h**

Nitrogen metabolism, Biological nitrogen fixation process by symbiotic bacteria and by free living bacteria and cyanobacteria, symbiotic and non-symbiotic nitrogen fixation. urea cycle, degradation and biosynthesis of essential and non-essential amino acids.

Reference

1. General Microbiology; Hans G. Schlegel, 7th Edition, Cambridge University Press M. Pelczar, E.C.S.Chan and N.R Krieg, Tata McGraw-Hill (2005)
2. Principles of Biochemistry by Nelson and Cox, 5thEdition, W.H. Freeman & Co., New York (2010)
3. Biochemistry; Donald Voet and Judith G Voet, John Wiley and Sons, Inc (2000).
4. Microbial Physiology; Albert G. Moat, 4th Edition, Wiley (2000)

Course code	Microbial diversity (HC)	Course type	L	T	P	C	CH
M21SO0202		HC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have basics of different classes of microorganisms and their importance.

Course Objectives:

1. Acquire knowledge and explore the diverse form microorganisms.
2. Illustrate the various importance's and applications of microorganism.
3. Explore the eukaryotic microorganisms and their applications in various field.
4. Explore the microbial ecology and its importance in environmental sustainability.

Course Outcomes:

By the end of the course the student will be able to:

1. Acquire the basic knowledge of diverse microbial life and their classifications.
2. Explore the recent methodologies in field of microbiology.

3. Exploit the biology and significance of bacteria and viruses in other areas.
4. Utilize the biological and significance of eukaryotic microbes for commercial purpose.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0202	CO1	3	3	2	3	2		2		1	3	3	3	2
	CO2	3	3	2	2	3		2		1	3	3	3	2
	CO3	3	3	2	2	3		2		1	3	3	3	2
	CO4	3	3	2	2	3		2		1	3	3	2	2

Course Content:

Unit-I Microbial World and Microbial Taxonomy 13 h

Origin and historical development of Microbiology, contribution of microbiologist, biogenesis & abiogenesis theory. Microbiology as interdisciplinary science, importance and scope. Five kingdom and domain classification, classification criteria in microbiology, Bergeys' classification, binomial nomenclature and classification systems in different microbial classes. Molecular tools of 16 S rRNA and 18S rRNA sequencing and phylogenetic analyses.

Unit-II Importance and Conservation of Microbial Diversity 13 h

Importance of microbial diversity in agriculture, forestry, environment, industrial & food biotechnology, animal & human health. Metagenomics approaches in the microbial community's determination. Importance of conservation. In situ conservation and Ex situ conservation. Role of culture collection centers in conservation.

Unit-III Bacterial and Viral Diversity 13 h

Brief history of bacteriology, general characteristics of bacterium, classification of bacteria based on the morphology, flagella and nutritional aspects, ultrastructure of bacterial cells and functions of bacterial structure.

Brief history of virology, general characteristics, classification of viruses, life cycle of bacteriophage (lambda, T4, M13), human virus (polio, hepatitis) and plant virus (TMV, CaMV) importance.

Unit-IV Fungal and Algal Diversity 13 h

Fungal characteristics, growth, reproduction, life cycle, classification & importance. Study of thallus structure, reproduction and life cycle of *Pythium, Rhizopus, Saccharomyces, Aspergillus, Penicillium, Fusarium and Agaricus*. Major applications of fungi in various fields.

Algae characteristics, structures, growth, reproduction, life cycle, classification & importance. Study of thallus structure, reproduction and life cycle of Cyanobacteria, Chlorella, Scenedesmus, Spirogyra, Diatoms and Gracilaria. Various applications of algae.

Reference

1. Introductory Mycology. Alexopoulos, C. J. and Mims, C. W. III edition, Wiley Eastern, New Delhi (2007).
2. Introduction to Modern Virology; Dimmock, N. J., Easton, A. J. and Leppard, K. N. 5th editionn. Blackwell publishing, USA (2001).
3. Natural Resource Conservation and Environment Management; Ghosh, A. Aph Publishing Corp. Calcutta (2003).
4. Brock Biology of microorganisms; Madigan M.T., Martinko M. J. and Parker, J. Pearson education (2003)
5. Microbiology; New Jercy. Pelczar, (Jr.) M. J., Chan, E. C. S. and Kreig, N. R; McGraw Hill, New York (2000)
6. Microbiology, Dynamics and Diversity. Perry, J.J. and Staley, J.T. 4th edn. Wesley Longman pub. New York (1997).
7. Microbial Diversity – Current Perspectives and Potential Applications. Satyanarayana, T. and Johri, B. N., I K Int. Pvt. Ltd. New Delhi (2005).

Course code	Bioinformatics and Biostatistics	Course type	L	T	P	C	CH
M21SO0203		HC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have basics of computers and mathematical calculations**.**

Course Objectives:

1. To introduce the basic knowledge of bioinformatics
2. To emphasis the applications of bioinformatics in biological research
3. To develop knowledge on various kinds of research design
4. To acquire basic knowledge on quantitative and qualitative analysis.

Course Outcome:

By the end of the course the student will be able to:

1. Apply the knowledge of computational biology in database analysis
2. Inculcate the fundamentals of binary genomic data analysis
3. Develop the new algorithms, methods and computational modeling in drug discovery
4. Inculcating application knowledge of biostatistical methods to evaluate biological data

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO3
M21SO0203	CO1	3	3	2	3	2		2		1	3	3	3	2
	CO2	3	3	2	2	3		2		1	3	3	3	2
	CO3	3	3	2	2	3		2		1	3	3	3	2
	CO4	3	3	2	2	3		2		1	3	3	2	2

Course Content:

Unit-I Biological databases and analysis **13 h**

Introduction to NCBI, NCBI, EMBL and other genomic data bases, Searching PubMed, BLAST, BLASTn, BLASTp, PSI-BLAST, searching sequence databases for sequence similarity; Multiple sequence alignment, Phylogenetic Analysis; Primer designing. Genome sequencing, sequencing platforms, sequence file types, data structures. Classification of microbes using genome sequencing. Differential gene expression prediction, functional annotation, functional enrichment analysis.

Unit-II Biological Data Retrieval and Analysis **13 h**

Introduction and scope of bioinformatics, biological information resources, genome sequence acquisition and analysis, data acquisition, biological databases, structure and annotation, data mining and data characteristics, scoring matrix, sequence alignment and database searches, pair wise and multiple sequence alignment.

Unit-III Bioinformatics in Drug discovery **13 h**

Conceptual model of protein structure, Structural types and conceptual models, Globular proteins, secondary structure, tertiary structure, integral membrane proteins and domains. Protein structure analysis, Molecular docking. Bioinformatics in the Pharmaceutical Industry- QSAR method; ADMET Predictions.

Unit-IV Data processing and Statistical analysis **13 h**

Processing and Analysis of Data; processing operations, problems in processing, Types of analysis, statistics in research, importance of statistical analysis. Measure of relationship- simple regression, multiple correlation and regression analysis, Analysis of variation (ANOVA) and co-variation, Meta-analysis, networking and Dataperl.

Reference

1. Introduction to Bioinformatics; Attwood, T. and P.S. David, Pearson Education Ltd., New York (2006)
2. Bioinformatics A Practical Guide to Analysis of Genes and Proteins., Baxevanis, AD., and Ouellette, BFF, 3rd Edition. John Wiley and Sons, New York (2006).
3. Bioinformatics and molecular evolution; Attwood TK. and Higgs, PG; Blackwell Publishers, London (2005).
4. Introduction to Bioinformatics; Lesk, AM. Oxford University Press, (2002).

Course code	Microbiology for Sustainable Environment	Course type	L	T	P	C	CH
M21SOS211		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should know about the role of microbes in environment and their importance.

Course Objective:

1. Facilitate the students to understand role of microbes in environment suitability.
2. Impart the knowledge of pollution control and waste management in society.
3. Utilization of beneficial microbes for sustainable agriculture.
4. Inculcate the knowledge to produce bioenergy from using waste.

Course Outcome:

By the end of the course the student will be able to:

1. Acquire the knowledge of role of various microorganisms in life cycle of organisms.
2. Identify the different types of pollution and their management strategies.
3. Increase the plant productivity using the ecofriendly microbial management in soil.
4. Impart the knowledge of energy production from microorganisms using organic waste.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SOS211	CO1	3	3	3	3		2	3		1	3	3	2	2
	CO2	3	2	2	2		2	3		1	3	3	2	2
	CO3	3	2	2	2			3		1	3	3	2	2
	CO4	3	3	2	2		2	2		1	3	3	2	2

Course Contents:

Unit-I Microbes in environment **13 h**

Role and diversity of microbes in nature, culture based and non-culture-based technique to study microbes in environment. Biogeochemical cycles-N, C, S and other cycles with the role of microorganisms. Pathogens of animals, humans and plants in nature and saprophytes in environment.

Unit-II Waste management **13 h**

Solid and Liquid waste Treatment of sewage and industrial effluents, Secondary waste treatment – aerobic, anaerobic and Composting, Biofuels – ethanol, methane and biohydrogen. Bioleaching of

Metals, Microbes as food, bioplastics from microorganisms, Genetically modified microbes – Application and hazards.

Unit-III Microbes in sustainable agriculture **13 h**

Role of Microbes in Agriculture – bioinoculants-types – bacterial, fungal and algal-nitrogen fixers, mineral solubilizes and other types, biodegradation of xenobiotic compounds-pesticides and petroleum products, techniques of bioremediation using microbes.

Unit-IV Microbes in waste to products **13 h**

Microbial Enhanced Oil Recovery, Bio-ethanol and bio-diesel production, commercial production from lignocellulosic waste, Algal biomass for fuel; Biogas production: Methane and hydrogen production using microbial culture.

References

1. Renewable Energy Environment and Development; Mahesh, Dayal; Konark Publishers(P) Ltd (1992).
2. Energy Technology; Rao S & Parulakar BB; Khanna Publishers, New Delhi (1994).
3. Wood and Cellulosic Chemistry; David N-S Hon DNS, Nobuo Shiraishi N; CRC Press, (2000).
4. Renewable Energy; Sorensen B, Academic Press (2010).
5. Sustainable Bioenergy and Bioproducts; Kasthurirangan G, van Leeuwen J, Robert C; Springer (2012).

Course code	Microbial food processing	Course type	L	T	P	C	CH
M21SOS212		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have knowledge of microorganism in food industries.

Course Objective:

1. To provide the basic knowledge of microbes associated with food.
2. To provide the impact of microbes in food spoilage and preservation.
3. To study the microbial process of beneficial microbes in food industry.
4. To acquire the knowledge of microbial food safety measures.

Course Outcome:

By the end of the course the student will be able to:

1. Better understanding of microbes associate with food.

2. Apply the scientific knowledge of evaluate the food spoilage and preservations.
3. Apply various microbial techniques in developing probiotics
4. Aware of various ethical reasoning in the discipline of food technology.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SOS212	CO1	3	3	3	3		2	3		1	3	3	2	2
	CO2	3	2	2	2		2	3		1	3	3	2	2
	CO3	3	2	2	2			3		1	3	3	2	2
	CO4	3	3	2	2		2	2		1	3	3	2	2

Course Content:

Unit I Microbial association of the food **13 h**

Structure, properties and metabolic functions of food constituents - composition and nutritive value of common food items - organoleptic properties of food– undesirable constituents in food. Microorganisms associated with food – Harmful and beneficial effects of microbes – Various factors viz. Intrinsic factors, extrinsic factors, Implicit factors, processing factors affecting microbial growth

Unit II Microbial food spoilage and food preservations **13 h**

Microorganisms and food spoilage– Spoilage of fruits, vegetables, fresh and processed meat, poultry and seafood and other miscellaneous foods - Food borne microbes causing health hazards- Food poisoning and intoxication - causes – symptoms – strategies employed for prevention and cure. Physical and chemical methods of preservation biopreservation, Food additives, definition, types and function, processing of selected dairy and poultry products- Methods for maintaining high quality food products

Unit III Probiotics and lactic acid bacteria **13 h**

Types of lactic acid bacteria – homofermentation and heterofermentation – Role of LAB in fermented food products – Genetic transfer systems in lactic acid bacteria -- Uncontrolled and controlled genetic alterations in lactic acid bacteria - Use of genetically modified LAB in food production - antibiotics – Probiotics and its importance.

Unit IV Microbial detection of Food and food laws **13 h**

Conventional methods - Membrane filters- Microscope colony counts - agar syringe methods - surface methods – rapid detection methods - nucleic acid-based methods - biosensors - immunological methods. Inspection – Microbial Indicators of product quality – Indicators of food safety – Microbiological safety of foods - control strategies – Hazard Analysis Critical Point System (HACCP concept)- Microbiological criteria.

Reference

1. Food Microbiology; James M. Jay, Martin J. Loessner, David A. Modern, 7th Edition, CBS Publishers, New Delhi, India (2011).
2. Food Oxidants and Antioxidants - Chemical, Biological, and Functional Properties; Grzegorz Bartosz, 1st Edition, CRC Press, New York, USA (2014).
3. Food Safety Management - A Practical Guide for the Food Industry; Yasmine Motarjemi, Huub Lelieveld, 1st Edition, Academic Press, London, UK (2014).
4. Food Microbiology; Adams, Martin R., Moss, Maurice O. 3rd Edition, Royal Society of Chemistry, Washington DC, USA (2017).

Course code	Microbial Metabolism and Physiology (LAB)	Course type	L	T	P	C	CH
M21SO0204		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should know the basics of biochemical testing and culturing of microbes.

Course Objective:

1. Explore the practical aspects of isolation and culturing of microorganisms
2. Exploit the practical knowledge of microbial enumerations.
3. Analytical techniques to evaluate the microorganisms associated with various samples
4. To identify and characterise the microbial products by various metabolic process.

Course Outcome:

By the end of the course the student will be able to:

1. Hands on experience in enumeration of microbes.
2. Development of practical skills in microbial technology.
3. Exploit the knowledge of microbial enzyme for commercial use.
4. Acquire enhanced microbial products to meet the nutritional requirements.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO0204	**CO1**	3	3	2	2	2	1	1	1	1	3	3	2	2
	CO2	3	2	2	2	2	1	1		2	2	3	2	2
	CO3	3	2	2	2	2		1		1	2	3	2	2
	CO4	3	2	2	2	1	1	2	1	1	2	3	2	2

Course Content:

1. Determination of viable and total number of cells.
2. Measurement of cell size, Growth – types of growth (synchronous, diauxic, batch),
3. Study factors affecting growth
4. Sporulation and spore germination in bacteria
5. Induction and repression of enzymes
6. Study of bacterial growth under aerobic, micro, aerophilic and anaerobic conditions
7. Morphological, Physiological and Biochemical tests of selected bacterial cultures.
8. Production of amino acids (Leucine and Glutamic acid) by microorganisms.
9. Production of vitamins (Vitamin-A) by microorganisms.
10. Production of extracellular enzymes-Amylase

Reference

1. Microbiology Applications – (A Laboratory Manual in General Microbiology), Benson H. J. Wm C Brown Publishers (2010).
2. A Laboratory Manual in Biochemistry; Cappuccino J.G. and Sherman N. Addison-Wesley (2010).
3. Laboratory Techniques in Biochemistry and Molecular Biology; Work T.S. and Work R.H.E. Elsevier Science (2010).

Course code	Microbial diversity LAB	Course type	L	T	P	C	CH
M21SO0205		**HC**	**4**	**0**	**0**	**4**	**4**

Prerequisites/Pre reading for the course:

Students should have theoretical knowledge of diverse class of microbes and their culturing aspects**.**

Course Objectives:

1. Enumeration of microflora from food and water samples.
2. Explore beneficial microflora from soil samples.

3. Identify the phytopathogens from soil microflora.
4. Quantitative estimation of microflora in various samples.

Course Outcome:

By the end of the course the student will be able to:

1. Inculcate the knowledge the of analysing the microflora in various samples.
2. Exploit the use of beneficial microflora for commercial purpose.
3. Identification and characterisation of phytopathogens.
4. Quantitative analysis of microflora in different samples

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO0205	CO1	3	3	2	2	2	2	1	1	1	3	3	2	2
	CO2	3	2	2	2	2	1	1		2	2	3	2	2
	CO3	3	2	2	2	2		1		1	2	3	2	2
	CO4	3	2	2	2	1	1	2	1	2	2	3	2	2

Course Content:

1. Isolation and identification of bacteria from food and water.
2. Isolation and identification and study of actinomycetes from soil.
3. Isolation and identification and study of cyanobacteria from soil / paddy field.
4. Isolation and study of bacteriophages from sewage.
5. Preparation of basic solid media agar slants and agar deep tubes for cultivation of fungi.
6. Isolation and identification of fungi from soil/cereals/water by serial dilution technique.
7. Staining of vesicular arbuscular mycorrhizae from soil.
8. Isolation and identification and study of algae from water.
9. Isolation and identification and study of phytopathogens.
10. Measurement of concentration of microorganism by hemocytometer

Reference

1. Microbiology; Presscott, L. M., Harley, J. P. and Klein, D. A., 4th edn. WCB Mc GrawHill, New Delhi (2000).
2. Microbial Diversity – Current Perspectives and Potential Applications; Satyanarayana, T. and Johri, B. N, I K Int. Pvt. Ltd. New Delhi (2005).
3. Biodiversity of microbial life; Stanley J.T, Reysenbach A; John Wiley 7 Sons Inc. Publication. New York (1997).

Course code	Bioinformatics and Biostatistics LAB	Course type	L	T	P	C	CH
M21SO0206		SC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have theoretical background of computers and software's for statistical studies.

Course Objectives:

1. To acquire fundamental knowledge of bioinformatics techniques.
2. Emphasis on application of bioinformatics in biological databases to resolve research problems.
3. Practical exposure to use wide applications computational biology and apply the same to resolve biological research.
4. To evaluate the biological data using the biostatistics tools.

Course Outcome:

By the end of the course the student will be able to:

1. Inculcate the knowledge of basic principles and concepts of computational biology.
2. Evaluate the basic information of biological database.
3. Conceptualize the molecular modelling of biological data and its applications.
4. Problem solving skills in qualitative and quantities analysis of data

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO0206	CO1	3	3	2	2	2	1	1	1	1	3	3	2	2
	CO2	3	2	2	2	2	1	1		2	2	3	2	2
	CO3	3	2	2	2	2		1		1	2	3	2	2
	CO4	3	2	2	2	1	1	2	1	1	2	3	2	2

Course Content:

1. BLAST-finding scores, E-values, Sequence alignment.
2. Phylogenetic analysis
3. Gene prediction methods
4. Primer designing
5. Protein structure prediction
6. Protein structure visualization and interpretations.
7. Protein Structure validation, active site predictions and molecular docking.
8. Developing a vector map using a software.
9. Calculation of mean, median and mode

10. Analysis of data for ANNOVA and student t-test using SPSS and GraphPad.

Reference

1. Introduction to Bioinformatics; Attwood, T., P.S. David. Pearson Education Ltd., New York (2006).
2. Bioinformatics A Practical Guide to Analysis of Genes and Proteins; Baxevanis, AD., Ouellette, BFF, 3rd Edition. John Wiley and Sons, New York (2006).
3. Bioinformatics and molecular evolution; Attwood TK, Higgs, PG; Blackwell Publishers, London (2005).
4. Introduction to Bioinformatics; Lesk, AM, Oxford University Press (2002).

THIRD SEMESTER

Course code	ORGANIC FARMING	Course type	L	T	P	C	CH
M21SLO301		OE	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should know the organic farming basics and its importance.

Course Objectives:

1. Understand the importance of organic farming in developing a sustainable agriculture system for ensuring adequate food production.
2. Acquire the holistic concept of organic farming as a self-sustainable unit in the ecosystem.
3. Get familiarized with the practices involved in the organic farming system.

Course Outcomes:

By the end of the course the student will be able to

1. Analyse the advantages of organic farming compared to conventional chemical agriculture.
2. Understand the significance of plant nutrient management following organic practices.
3. Apply the knowledge gathered in implementing the practices in organic crop production and plant protection.
4. Explain the basic principles involved in organic farming production, certification and marketing.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SLO301	CO1	2	3		3	2		3				3	3	2

	CO2	3	2									2	2	3
	CO3	3	2		3			3		3	3	3	3	2
	CO4	2	3	3	3			3				3	3	3

Course Content:

Unit I Introduction to Organic Farming 13 h

Organic farming, concept and development of organic farming. Principles of organic farming & need for organic farming, Agencies and institutions related to organic agriculture. Farm components for an organic farm. Benefits of organic farming. Conventional farming v/s organic farming. Scope and present state of organic farming; its relevance to India and global agriculture and future prospects.

Unit II Organic Plant Nutrient Management 13 h

Organic farming systems: Soil tillage, Choice of Varieties, crop rotation, multiple cropping systems, intercropping. Propagation: planting materials and seed treatments. Water management, Organic manures: Green manuring, Composting: Composting methods, Vermicomposting, Organic amendments and sludges, biogas. Bio-fertilizers-: methods of application and advantages.

Unit III Organic Plant Protection 13 h

Plant protection: cultural, mechanical methods. Biological methods: botanical pesticides, biopesticide, biocontrol agents. Weed management. National and international Standards for organic inputs- plant protection; Integrated pest management. Organic crop production methods: arecanut, okra. Live-stock component and management in organic farming.

Unit IV Organic Certification 13 h

Farm economy; Basic concept of economics- Demand, supply, Economic Viability of a farm. Basic production principles, Reducing expenses, ways to increase returns. Cost of production system. Marketing, Imports and exports. Policies and incentives of organic production, Farm inspection and certification: Income generation activities: Apiculture, Mushroom production, Terrace farming.

Reference

1. Organic Farming – Theory and Practice. Palaniappan, S.P and Anandurai, K; Scientific Publ. Rao (1999).
2. Small Farmer Focused Integrated Rural Development: Socio-economic Environment and Legal Perspective: Parisaraprajna Parishtana, Bangalore, Publ.3, B.V.V. (1995)
3. Soil Organisms and Litter Decomposition in the Tropics. Reddy M.V. Oxford & IBH (1995)
4. Hand Book of Organic Farming, Sharma, A; Agrobios. (2002).
5. Technology for Production of Natural Enemies, Singh, S. P. PDBC, Bangalore, (1994)
6. Soil Microbiology, Subba Rao, N.S.; Oxford & IBH 2002.

Course code	Nanobiology and Space Microbiology	Course type	L	T	P	C	CH
M213SOS311		OE	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have a knowledge of nanotechnology in life sciences and its importance.

Course Objectives:

1. Better understand of the synthesizes and characterization of nanomaterials
2. Describe various types of nanomaterials and their applications
3. Apply the knowledge to understand the application of nanomaterials in allied sciences
4. Have Insight and deep knowledge of space microbiology and its importance

Course Outcomes:

By the end of the course the student will be able to

1. Describe the basic science behind the properties of materials at the nanometre scale
2. Understand the principles behind advanced experimental and computational techniques for studying nanomaterials.
3. Systematically solve scientific problems related specifically to nanotechnological materials using conventional scientific and mathematical notation.
4. Explain primary aspects of space microbiology that have been studied to date.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M213SOS311	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Contents:

Unit I Concepts, synthesis and characterization of nanomaterials 13 h

Basic and functional principles of nanotechnology, basic biology principles and practice of micro and nanofabrication techniques. Synthesis approaches and methods, characterization of nanomaterials by spectroscopic and microscopic techniques, Biological production of nanoparticles, macro molecular assemblies and its significances.

Unit II Various nanostructures and their applications **13 h**

Bacterial structure relevant to nano-biotechnology, Cubosomes, Dendrimers, DNA Nanoparticle Conjugates, DNA Octahedron, Fullerenes, Nanoshells, Carbon Nanotubes, Nanopores, Nano structured Sillicon, Viruses as nano-particles, nano chemicals and application. Nanomaterials used in various fields and their importance.

Unit III Allied applications of Nanobiotechnology **13 h**

Drug delivery tools through nanotechnology, tumor targeting and other diagnostic applications, nanoparticle-based immobilization assays, quantum dots technology and its application, immuno-nanotechnology. Biosensors and nano-biotechnology, principles used in construction of microelectronic devices, sensors and macro mechanical structures and their functioning. DNA based Nanostructures- DNA-protein nanostructures-Methods-Self assembled DNA nanotubes-Nucleic acid Nanoparticles, DNA as a Biomolecular template-DNA branching Metallization-Properties.

Unit IV Space Microbiology **13 h**

Space Microbiology: An Overview, Monitoring of astronauts microbial flora: Alterations in the load of medically important microorganisms, ESA STONE experiment. Evaluating the Biological Potential in Samples Returned from Planetary Satellites and Small Solar System Bodies.

References

1. Nanobiotechnology- concepts, applications and perspectives, Niemeyer, Christof m. Mirkin, Chad A., Wiley publishers (2000)
2. Nanobiotechnology of biomimetic membranes, Martin, Donald (edt), Springer Verlag publishers, (2000).
3. Nanotechnology in Drug Delivery; MelgardtM.deVilliers, PornanongAramwit, Glen S.Kwon, Springer-American Association of Pharmaceutical Scientists Press, (2009).
4. Bio-Nanotechnology; Elisabeth S.Pappazoglou, Aravind Parthasarathy, New Age publishers, (2010).
5. Biomedical Nanostructures: Kenneth E.Goonsalves, Craig R.Halberstadt, Cate T. Laurecin, Lakshmi S.Nair, Springer, (2011).

Course code	Marine Microbiology	Course type	L	T	P	C	CH
M213SOS312		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have knowledge of aquatic microbiology and their importance.

Course Objectives:

1. Explain the scope and challenges in the field of marine biotechnology

2. Develop better knowledge about marine ecosystems, biodiversity & taxonomy, tools and techniques used and role of marine organisms in biogeochemical cycles
3. Realize the biotechnological importance and to explore the potential of marine organisms for human betterment
4. Demonstrate the various techniques and tools necessary for studying marine microbial diversity and its applications.

Course Outcomes:

By the end of the course the student will be able to

1. Develop knowledge about marine biotechnology, compare the various marine ecosystems and explain the tools and techniques used for sample collection, isolation of micro and macro-organisms and to study their taxonomy.
2. Analyze the importance of marine organisms in various marine pathogenic microbes and their transmission to land
3. Build knowledge on process of drug discovery from marine organisms and various assays and techniques related to it
4. Utilize marine organisms for food, fuel, agriculture, environment, cosmetics, industrial feedstock etc.,

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M123SOS312	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

Unit I Marine Ecology and Diversity **13 h**

Benthic and Pelagic Zone; Photic, dysphotic and aphotic zones- importance and their significance. Biological divisions of the sea- estuaries and backwaters, lagoons, mangroves, coastal waters, inshore, offshore, deep sea/oceanic zone. Sampling, cultivation and taxonomy of microorganisms and metagenomic approaches. Bacteria, fungi, algae and archaea, Extremophilic microorganisms.

Unit II Marine microbial pathogen **13 h**

Microbial pathogens in marine environment - diversity, sources and detection of pathogens in recreational water, impact of harmful algal blooms, microbial pathogens of seafood. Effect of climatic changes in marine ecosystems.

Unit III Marine Pharmacology **13 h**

Marine derived drugs in preclinical and clinical trials- FDA and EMEA approved marine derived drugs, their use and mode of action. Screening of drugs High-throughput Screening Assays (HTS), Enzyme inhibitory assays, cytotoxicity assay; antimicrobial assay; DNA laddering assay; Apoptosis assay and screening for biologically active molecules from marine microorganisms.

Unit IV Marine microbes for bioprospecting **13 h**

Marine organisms for Biofuels and bioenergy, Bioremediation, Biofouling, Biosurfactants. Marine natural products as cosmetics-cosmeceuticals, algotherapy; Thalassotherapy; Enzymes; food, supplement, nutrition and energy drinks. Marine algae as fish feed, manure and fertilizers. Recent developments in marine microbiology.

References

1. Aquaculture Microbiology and Biotechnology; Didier Montet, Ramesh C. Ray (Eds.), Volume Two 1st Edition, Science Publishers, (2011).
2. Introduction to Marine Biology; George Karleskint, Richard Turner, and James Small (Eds.) 4th edition, Brooks Cole Publishers, (2013).
3. Bioactive Marine Natural Products; Dewan S. Bhakuni and, D.S. Rawat (Eds.), Springer publishers, (2010).
4. Marine Microbiology: Ecology and Applications; Munn and Munn BIOS, Scientific Publisher, (2011).

Specialization in Industrial & food Microbiology

ourse code	Food Microbiology	Course type	L	T	P	C	CH
M21SO03Z01		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have basis of microbial food processing and preservation aspects.

Course Objectives:

1. Understand the principles of microorganisms during various food-processing and preservation steps.
2. Isolation, identification, and enumeration of the most common microorganisms found in specific food products.
3. Recognize specific types of microbial spoilage during various food shelf-life stages.
4. Analyze different foods for presence of hazardous microorganisms using traditional and modern food microbiology technology.

Course Outcomes:

By the end of the course the student will be able to

1. Better understand the factors and sources of food contaminants
2. Describe the situations where improper food handling and storage may lead to the spoilage or contamination of food.
3. Identify desirable microorganisms and their effects in preservation and fermentation.
4. Develop the beneficial microbes in health and other allied food industries.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z01	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

Unit-I History and development of Food microbiology 13 h

History of Microorganisms in Food-developments Common Food borne Bacteria, Role of Molds, and Significance of Microorganisms in Foods. Parameters Affecting Microbial Growth: Intrinsic, Extrinsic. Combined Intrinsic and Extrinsic Parameters-lactic antagonism and hurdle concept.

Unit-II Microorganisms in Foods and methods for detection 13 h

Fresh meat, Processed meat and poultry, Culture, Microscopic, and Sampling Method for detecting microbes, Physical, Chemical methods, Whole animal assays, Immunological methods. Microbial Food Spoilage and Food borne diseases-*Staphylococcal, E coli*, Salmonellosis, shigellosis, Listerial infections. Mycotoxins, Aflatoxins Alternaria Toxins, Toxigenic Phytoplankton's and viruses.

Unit-III Food Preservation & Principles of Quality Control 13 h

Chemicals, antibiotics, Radiation, Low and high temperature, High-Pressure Processing Pulsed Electric Fields. Aseptic Packaging, Manothermo, sonication, Microbiological quality standards of food, Government regulatory practices and policies, FDA, HACCP and ISI.

Unit -IV Applications of Food Microbiology 13 h

Applications of Food Microbiology: Beneficial uses of microorganisms in Food Intestinal Beneficial Bacteria-Concept of Prebiotics and Probiotics, Genetically modified foods. Biosensors in foods.

References

1. Food Microbiology; 2nd Edition, Adams, New Age Publishers, (2011).
2. Basic Food Microbiology; Banwart George J.; Elsevier, (2010).
3. Food Microbiology: Fundamentals and Frontiers; Dolle; Springer, (2010).
4. Fundamentals of Dairy Microbiology; Prajapati, New Age Publishers, (2011).
5. Essentials of Food Microbiology; John Garbult. Arnold International Students Edition, (2011).
6. Microbiology of Fermented Foods; Volume II Brian J. Wood, Elsiever Applied Science Publication, (2011).

Course code	Food Chemistry and processing	Course type	L	T	P	C	CH
M21SO03Z02		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have knowledge macro and micronutrients present in the foods.

Course Objectives:

1. Scientific basis of water and carbohydrates in nature and their interactions.
2. Better understanding of chemistry and importance of the oils and fats.
3. Gain a knowledge about the food processing and preservation technology
4. Understand the various food packaging technology and its mechanisms.

Course Outcome:

By the end of the course the student will be able to

1. Explain the scientific basis of the role of water and carbohydrates with their interactions.
2. Understand the chemistry of oils and fats with its importance.
3. Describe the various food processing and food preservation of different food items.
4. Explore the various packaging materials and process involved in food packing.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PS O3
M21SO03Z02	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

Unit I Water and Carbohydrates for food **13 h**

Role of water and chemistry of carbohydrates and proteins, Physicochemical properties of water, water weak interactions in Aqueous Systems. Role of water in food, Moisture in foods and determination of moisture. Sources of Carbohydrates and proteins, composition and factors

affecting composition of foods, Chemistry of cellulose, starches, other polysaccharides - starch enzymes, Gel formation and starch degradation - Pectic substances, their occurrence, structure, properties and applications in foods., Classification of proteins, physical and chemical properties of proteins, Conformation, functional properties of proteins in foods, hydrolysis of proteins - Changes in proteins during processing - Determination of Proteins.

Unit II Chemistry of Oils and Fats **13 h**

Nomenclature and classification, Physical and chemical properties of fats, rancidity and flavour reversion, processing of oil-bearing materials, refining of oils and fats, fat hydrolysis and trans-esterfication, hydrogenation, Changes in Acyl lipids of foods, Classification and Properties of Unsaponifiable constituents. shortenings and spreads - Emulsions, Definition, surface activity, surface film theory of emulsions, properties and types of emulsions, emulsifying agents, their chemistry during processing - Essential oils, Chemistry of occurrence, Extraction - Terpene oils and their use in foods.

Unit III Food processing and preservations **13 h**

Food processing, preservation Processing and preservation by drying, methods of food concentration: freeze concentration, Ultra-filtration, reverse osmosis, evaporation-Drying, low Temperature, dehydration of fruits, vegetables, milk and animal products. Various methods employed in production of dehydrated commercial products, selection of methods based on characteristics of foods to be produced, advantages and disadvantages. Food irradiation, history and mechanism, the electro-magnetic spectrum, forms of radiant energy. Principles of using electromagnetic radiation in food processing with advantages and disadvantages. Processing and preservation by non-thermal methods. Food additives; Definition, types and functions and safety aspects. Chemical and metal preservatives- type I and type II and permissible limits.

Unit IV Food packaging **13 h**

Objectives and functions of packaging and packaging materials; Types and selection of packaging materials: Paper: pulping, fibrillation and beating, types of papers; Glass: composition, properties, types of closures; Metals as packaging materials: Tinplate containers, tin free steel (TFS), aluminum containers, lacquers; Plastics: types of plastic films, laminated plastic materials, co-extrusion, edible films, biodegradable plastics. Aseptic conditions during packaging.

Reference Books

1. Food Science and experimental foods; Swaminathan N; Ganesh Publications, Madras, (1987).
2. Food chemistry; Meyer L.M. Van Noustrand Reinhold co., New York. 34 (1969).
3. Foundations of Food Preparation; Peckham, C, The Macmillan co., London (1979).
4. Food Theory and Applications; Paul P.C, Palmer H.H John wiley and Sons, New York (1972).
5. Introductory foods; Bennion M, Hughes, D; Macmillan publishing Co., New York (1975).

Course code	Bioprocess Technology	Course type	L	T	P	C	CH
M21SO03Z03		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have knowledge about the fermentation and its products from microbes.

Course Objectives:

1. Gain a knowledge of the basis for upstream process in fermentation technology.
2. Better understand of working principle and design of the bioreactor.
3. Optimize the techniques and formulate the downstream processing of the products.
4. Design the plan for production of industrially important products through fermentation.

Course Outcomes:

By the end of the course the student will be able to

1. Describe the largescale cultivations of microbes for product development.
2. Comprehend the design of microorganisms for fermentation process.
3. Standardize the optimum techniques for the maximum production and separation of fermented products.
4. Apply the knowledge of fermentation for various products from microorganisms.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PS O3
M21SO03Z03	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

UNIT I Introduction to Bioprocess 13 h

Engineering: Basic concepts of bioprocess engineering- A brief survey of organisms, processes, products. Fermentation- Bacterial, Fungal and Yeast, Biochemistry of fermentation. Industrial strain improvement for better productivity; Fermentation media and Fermentation Process: Natural and synthetic media; Strategies for media formulation, sources of carbon, nitrogen, vitamins and minerals with anti-nutritional factors. Sterilization- Dry and moist heat; Types of fermentation process- submerged, surface and solid state.

UNIT II Bioreactors & fermentation process 13 h

Architecture of advanced bioreactors and their working mechanisms; Design features; Heat and Mass transfer; Specialized bioreactors- design and their functions; Bioreactors- Airlift, Tubular, Membrane, Tower, Fluidized-bed and Packed-bed; photo bioreactors and disposable reactors. Modes of cultivation- batch, fed- batch and continuous fermentation; Kinetics of fermentation, bioprocess control, monitoring of variables- Dissolved oxygen (DO), temperature, agitation, pH and pressure

UNIT III Downstream processing 13 h

Overview of unit operations and their principles, Physical and rheological characteristics of fermentation broths; Pre-treatment-Cell disruption, heating and chemical treatment; solid- liquid separation- filtration and centrifugation; Product isolation- Adsorption, precipitation and extraction; Purification- Chromatography- Size exclusion, affinity and ion-exchange and HPLC; Finishing operations – Freeze-drying, drying and crystallization.

UNIT IV Microbial fermentation & products 13 h

Microbiology of cheese and beverage fermentation. Microbiology of fermented milk products (acidophilus milk, yoghurt). Role of microorganisms in beverages – tea and coffee fermentations. Vinegar Fermentation Biosensors in food, Applications of microbial enzymes in dairy industry [Protease, Lipases]. Utilization and disposal of dairy by-product - whey. Production of value-added products: Bio preservatives, Biopolymers, Industrial Enzymes, Bio fuels, Cheese, Beer and Single Cell Protein. Fermented meat products. Production of recombinant proteins having therapeutic and diagnostic applications, and vaccines.

Reference Books:

1. Biotechnology; Satyanarayana, U, Books and Allied (P) Ltd (2005).
2. A Textbook on Biotechnology; Kumar, H.D. 2nd Edition. Affiliated East West Press Pvt. Ltd (1998).
3. Concepts in Biotechnology; Balasubramanian, D. et al., Universities Press Pvt. Ltd., (2004).
4. Basic Biotechnology; Ratledge, Colin and Bjorn Kristiansen 2nd Edition Cambridge University Press (2001).
5. A Textbook of Biotechnology; Dubey, R.C. S. Chand and Co. Ltd (2006).

Course code	Food Microbiology Lab	Course type	L	T	P	C	CH
M21SO03Z04		SC	0	0	4	2	4

Prerequisites/Prereading for the course:

Students should have theoretical knowledge of fermented food products and preservation techniques.

Course Objectives:

1. Better understanding of the laboratory design in food safety laboratory.
2. Hands on experience in isolation and identification of microorganisms from various food items.
3. Detection and quantification of microorganisms in food samples.
4. Modern molecular detection technology used in screening the microbes in food samples.

Course Outcomes:

By the end of the course the student will be able to

1. Deep understanding of the laboratory design of food testing laboratory and its importance.
2. Practical handling of the isolation and identification of the microorganisms testing of the food items.
3. Acquire various culture-based methods for detection and quantification of microbes in food samples.
4. Apply modern molecular based methods for detection and quantification microbes in food.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO03Z04	CO1	2	1	1	2	1	1	1	1	1	3	2	1	1
	CO2	1	2	2	1	2	1	1		2	2	1	1	1
	CO3	1	2	2	1	1		1		1	2	1	1	2
	CO4	1	1	2	2	1	1	2	1	1	2	2	2	1

Course Content:

1. Introduction to HACCP and food safety conditions in industry.
2. Sterility tests - Direct inoculation, direct immerse inoculation & membrane filtration.
3. Isolation and identification of microbes from different food source-Canned foods, Meat, Vegetables, Fruits & Milk
4. Microbial limit test & Anaerobic plate count
5. Determination of enteric microorganisms (Petrifilm/VRBA)-Yeast and moulds petrifilm
6. Determination of MRSA from food
7. Bacteriological examination of water by multiple tube fermentation test (MPN)
8. Microbial characterizations techniques-Determination of high osmotic conditions bacteria, Determination of lactic acid producing microbes, sulphate reducing bacteria, nitrifying bacteria, Oxidase listeria rapid test, AOAC Disinfectant test, Koch's postulates for bacterial and fungal pathogen.
9. Determination of toxic substances from food, Preservative efficiency test, MIC and MLC. Bio-burden tests, Organoleptic tests
10. Determination of microbes by PCR

Reference:

1. A Textbook on Biotechnology; Kumar, H.D 2nd Edition. Affiliated East West Press Pvt. Ltd (1998).
2. Concepts in Biotechnology; Balasubramanian, D. et al., Universities Press Pvt. Ltd (2004).
3. Basic Biotechnology; Ratledge, Colin and Bjorn Kristiansen 2nd Edition Cambridge University Press (2001).

Course code	Food chemistry LAB	Course type	L	T	P	C	CH
M21SO03Z05		SC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have basic details of food quality and testing requirements.

Course Objective:

1. Gain a knowledge of the various equipment and labware used in the food testing laboratory.
2. Analyze the macro and micronutrients and their roles in the food preparations.
3. Determine the physicochemical properties of the food and their preparations.
4. Qualitative and quantitative analysis of the metabolites in food.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z05	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Outcome:

By the end of the course the student will be able to

1. Describe and design the food testing laboratory and its requirements.
2. Apply the practical quantification of macro and micronutrients in foods.
3. Exploit the physicochemical properties of the food items
4. Analyze the various metabolites in the food and their preparations.

Course Content:

1. Introduction to equipment and labware used food analysis laboratory**.**
2. Analysis of Macronutrients in food-Determination of fat content, water content, vitamins C and cholesterol
3. Analysis of Micronutrients from food - Sulphate, calcium, phosphates, magnesium, tannins

4. Determination of moisture content, alkalinity and salt
5. Analysis of water-Physical test and chemical test
6. Analysis of milk - Organoleptic test, litmus test, Methylene blue dye reduction test, alcohol test, Adulterants and preservatives, ALP activity
7. Analysis of fruits and vegetables - Determination of salt content, acidity, added sugar and Peroxide activity in fruit content.
8. Analysis of oils and fats-Determination of specific gravity & Polybromide test for mustered oil
9. Determination of food additives - Colour (saffron, curcumin), Flavours, Hemictant (sorbitol), Emulsifiers (lecithin, mono glycerides) and Preservatives (Benzoic acid)
10. Extraction and analysis of secondary metabolites from plants.

References

1. Concepts in Biotechnology; Balasubramanian, D. et al., Universities Press Pvt. Ltd (2004).
2. Basic Biotechnology; Ratledge, Colin and Bjorn Kristiansen 2nd Edition Cambridge University Press (2001).

Course code	Bioprocess Technology LAB	Course type	L	T	P	C	CH
M21SO03Z06		SC	0	0	0	2	4

Prerequisites/Pre reading for the course:

Students should have theoretical knowledge of fermentation and their products.

Course Objective:

1. Develop skills and competencies in the fermentation process.
2. Practical skills on the downstream processing in the products.
3. Develop various skills to purify the fermented products.
4. Qualitative and quantitative analysis of the various fermented products.

Course Outcome:

By the end of the course the student will be able to

1. Analyze the various skills in extraction of products from fermentation.
2. Apply the practical skills in purification and analyze the products.

3. Exploit the various methods in characterization of the products.
4. Demonstrate the various process of characterization of the products.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO03Z06	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

1. Extraction of enzyme from plant or microbial sources
2. Downstream processes - Crude extraction of enzyme
3. Purification of crude enzyme-Precipitation of enzymes (salt/solvent) and Dialysis
4. Purification by chromatography 0Ion exchange and Gel filtration chromatography
5. Enzyme assay - Total protein estimation by Biuret method and 280 methods
6. Determination of specific activity
7. SDS PAGE
8. Native PAGE and Zymography
9. Immobilization of Enzymes and formulation
10. Production of lactic acid and critic acid and their estimation

References

1. Concepts in Biotechnology; Balasubramanian, D. et al., Universities Press Pvt. Ltd (2004).
2. Basic Biotechnology; Ratledge, Colin and Bjorn Kristiansen 2nd Edition Cambridge University Press (2001).

Specialization in Agriculture & Environmental Microbiology

Course code	SOIL MICROBIOLOGY	Course type	L	T	P	C	CH
M21SO03Z07		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have important roles of microorganisms in the environment and agriculture.

Course Objective:

1. To inculcate student fundamentals of soil microbiology
2. To understand the role of microbes in soil nutrition management
3. To explore the interaction of soil microbiota
4. To exploit the beneficial microflora to increase the fertility of the soil and bioremediation

Course Outcome:

By the end of the course the student will be able to:

1. Acquire the basic knowledge soil microflora.
2. Understand the implications of microflora in biogeochemical cycles.
3. Interpret interaction of microflora in soil ecosystem.
4. Exploit the knowledge of microbes to increase soil fertility and bioremediation.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z07	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

UNIT I Soil microbes **13 h**

Discoveries in soil Microbiology; distribution of microorganisms in soil, Autochthonous, Allochthonous and Zymogenous microbes, quantitative estimation of microorganisms in soil, role of microorganisms in soil fertility; influence of soil and environmental factors on micro flora, moisture, pH, temperature, organic matter, agronomic practices and Soil health card.

UNIT II Biogeochemical cycles **13 h**

Biogeochemical cycles, Carbon cycle, organic matter decomposition, humus formation, Nitrogen cycle - nitrogen fixation, ammonification, nitrification, denitrification, reactions – organisms involved. Nitrogen fixation – symbiotic - root nodulation, non-symbiotic, associative organisms, nitrogenase, hydrogenase, nif gene, nod gene. Microbial transformation of phosphorus, solubilization by phosphobacteria and P- mobilization by mycorrhizal fungi, Microbial transformation of sulphur - sulphur toxicity and sulphur bacteria.

UNIT III Interaction of microbes with soil ecosystem **13 h**

Interaction between soil microbes–Neutralism, Commensalism, Symbiosis, Synergism, Amensalism, Parasitism, Predation and Competition. Interrelationships between soil microbes and plants, Rhizosphere concept, R:S ratio, rhizoplane; spermosphere; phyllosphere, Mycorrhizae-types, Rumen flora, Insects microbial interactions.

UNIT IV Biofertilizer and bioremediation **13 h**

Biofertilizers – *Rhizobium, Azotobacter, Cyanobacteria, Azolla*, and VAM. Biopesticides – *Bacillus thuringiensis, Psuedomonas fluroscence* - 84, *Trichoderma viridae, Bavaria bassiana*, Nuclear Polyheadrosis Virus. Biodegradation – Cellulose, Lignin. Biodetoriation – Wool, Leather. Bioleaching- Copper, Uranium. Biomagnification, Bioremidiation – Degradation of DDT, Atrazine (Xenobiotic Compounds) and Cleanup oil spills – *P. putida*.

REFERENCE BOOKS:

1. Soil Microbiology; Subba Rao NS. Fourth edition, Oxford and IBH Publishing Co.Pvt. Ltd., New Delhi (2004).

2. Soil Microbiology; Mishra RR First edition, CBS Publishers and distributors, New Delhi (2004).
3. Disease of Crop Plants in India; Rangaswami G and Mahadevan A, Fourth edition, PHI Learning (P) Ltd., New Delhi (2002).
4. Agricultural Microbiology; Rangaswami G and Bagyaraj, Second edition, PHI Learning (P) Ltd., New Delhi (2002).
5. Soil Microbiology; Robert, L Tate First edition, John Wiley and Sons, Inc. New York (1995).
6. Microbial Ecology; R,M, Atlus and Richard Bartha Fourth edition, An imprint of Addison Wesley Longman, Inc, New York (2000).

Course code	PLANT PATHOLOGY	Course type	L	T	P	C	CH
M21SO03Z08		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have knowledge of plant disease caused by microorganisms.

Course Objective:

1. Understand the fundamentals of plant pathology.
2. Exploit the role of pathogens in disease development.
3. Familiarize with the pathogen interactions with its host.
4. Better understanding of the host defense mechanism against plant pathogens.

Course Outcome:

By the end of the course the student will be able to:

1. Identification of phytopathogens prevailing conditions.
2. Explore the basic knowledge of plant pathogens biology.
3. Comprehend and explain the host-parasite interaction.
4. Design the defence mechanism against plant pathogens to control diseases.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3

M21SO0Z08	**CO1**	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Contents:

UNITI Fundamentals of plant pathology **13 h**

Importance and history, definitions and concepts of plant disease. Classification of Plant Diseases. Classification of fungi & bacteria diseases. Phytopathology – Classification of plant diseases, signs, and related terminology. Bacterial disease – Citrus canker, Blight of paddy, Fungal Disease- Red rot of sugarcane, Black stem rust of wheat, Tikka leaf spot, Wilt of cotton, Viral Disease – TMV, Vein clearing disease. Principles and methods of plant disease management, integrated plant disease management. Plant nematodes and its managements.

UNIT II Plant pathobiology **13 h**

Study of plant pathogens and phanerogamic plant parasites. General characters, reproduction, survival and dispersal of important plant pathogens, and symptomatology.

UNIT III Plant-pathogens interactions **13 h**

Host parasite interaction, recognition concept and infection, role of environment and host nutrition on disease development. Disease development-role of enzymes, toxins, growth regulators.

UNIT IV Plant Defence mechanism **13 h**

Defence mechanisms - oxidative burst; Phenolics, Phytoalexins, PR proteins, Elicitors. Altered plant metabolism as affected by plant pathogens. Genetics of resistance; ëRígenes; mechanism of genetic variation in pathogens; molecular basis for resistance; marker-assisted selection; genetic engineering for disease resistance. MiRNA and plant defense mechanism.

REFFERENCES

1. Plant Pathology; Agrios G N. 5th Ed. Academic Press, New York (2005).
2. Physiological Plant Pathology; Heitefuss R, Williams PH. Springer Verlag (1976).
3. Plant Pathology; Mehrotra R S, Aggarwal A. 2nd Ed. Oxford &IBH, New Delhi (2003).
4. Introduction to Principles of Plant Pathology; Singh R S. Oxford & IBH, New Delhi (2002).

5. Disease and Insect Resistance in Plants; Singh DP, Singh A. Oxford &IBH, New Delhi (2007).

Course code	Biomass and energy systems	Course type	L	T	P	C	CH
M21SO03Z10		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should know about the role of microorganisms as important bioresources.

Course Objective:

1. Understanding of various renewable feedstocks, their availability and attributes for biofuels production.
2. Thorough understanding of the broad concept of generations of biofuel production from biomass.
3. Explore the other low-cost Agri-residues, biowastes, anaerobic digestion and biodiesel production.
4. Analyze and design processes for biofuel production using microbes.

Course Outcome:

By the end of the course the student will be able to

1. Identify and apply potential biomass feedstocks including energy crops.
2. Understand the existing and emerging biomass to energy technologies.
3. Develop a critical thinking about sustainability &resilience.
4. Determine potential solutions for energy needs and problems by incorporating the bioenergy technologies being explored.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO03Z10	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

Unit-I Biomass **13 h**

Fundamentals of bioenergy/biofuel; terms and concepts, origin, characteristics, advantages, and dis-advantages, use and cost of different types of biomass resources (renewable feed stocks): agricultural energy crops, agro-horticultural Lignocellulosic residual material and other biogenous waste-production, availability and attributes for bioenergy production.

Unit II Bioenergy production for sustainable environment **13 h**

General principles of the carbon cycle, greenhouse effect and global climate change. Bioeconomy and sustainable bioenergy system, Current and projected future technologies for producing biofuels such as ethanol, biodiesel from oil crops, microbial fuel cells, biohydrogen.

Unit-III Biofuel production **13 h**

Biofuel generations, Pre-treatment technologies, structure and function of lignocellulosic biopolymers, various types of pre-treatment technologies (Physical, mechanical, chemical, biochemical, ionic liquids etcetera) bioconversion of biomass to biofuel; concept of pseudo-lignin and inhibitors, biodiesel production; environmental impacts of biofuel production; concept of Biorefinery, value-added product generation in an integrated approach, processing of biofuel residues-case studies on combined heat and power (CHP)generation. The role of transgenic plants and algae.

Unit- IV Bioenergy Technology **13 h**

Anaerobic digestion process for biogas production, Inoculum-its stability and methane potential, Process microbiology, role of microbes, types and characterization, Effect of pH, temperature, nutrients, organic load ingrate (OLR) and hydraulic retention time (HRT) on biogas production from biogenic waste, Storage and stability of digestate-health and safety issues, Up-gradation of biogas to methane. Life cycle assessment of biofuels and biofuel technologies, India's energy demand and supply management, energy cropping, energy needs for the future: regional prospects.

References

1. Renewable Energy Environment and Development; Mahesh & Dayal; Konark Publishers(P) Ltd (1992).
2. Energy Technology; Rao S & Parulakar BB Khanna Publishers, New Delhi (1994).
3. Wood and Cellulosic Chemistry; David N-S Hon DNS & Nobuo Shiraishi N. CRC Press (2000).
4. Renewable Energy; Sorensen B, Academic Press (2010).
5. Sustainable Bioenergy and Bioproducts, Kasthurirangan G, van Leeuwen J, Robert C. Springer (2012).

Course code	Soil Microbiology Lab	Course type	L	T	P	C	CH
M21SO03Z10		SC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have knowledge of microbes and their important role in agriculture.

Course Objective:

1. To explore the aspects of microbial culture techniques
2. Formulation and production of biofertilizers
3. To develop mass production of biofertilizers
4. To understand the effect of biofertilizer in the growth of plants

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO03Z10	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Outcome:

By the end of the course the student will be able to

1. Apply the knowledge of microbial enumeration and culture techniques.

2. Formulate of liquid and solid biofertilizers
3. Mass production of biofertilizers through industrial fermentations.
4. Evaluate the quality and growth parameters of biofertilizers.

Course Content:

1. Mother culture preparation and preservation techniques
2. Isolation and purification of phosphate solubilizing microorganisms in soil.
3. Mass production of carrier-based formulation of *Azospirillum brasilense, Pseudomonas fluorescens* and *Frateuria aurantia*
4. Industrial mass production of liquid formulation of *Azospirillum spp.*
5. Developing liquid formulations of bio fertilizers to increase shelf life
6. Hands on training of operation of industrial fermenters
7. Concentration of bacterial suspension through centrifugation and spray drying
8. Quality analysis of bio-fertilizers with respect to FCO standards
9. Evaluating effect of bio-fertilizers on crop growth in pot study experiment
10. Bio-fertilizer consortium preparation.

References

1. Agricultural Microbiology; Rangaswami G and Bagyaraj, Second edition, PHI Learning (P) Ltd., New Delhi (2002).
2. Soil Microbiology; Robert, L Tate First edition, John Wiley and Sons, Inc. New York (1995).
3. Microbial Ecology; R,M, Atlus and Richard Bartha Fourth edition, An imprint of Addison Wesley Longman, Inc, New York (2000).

Course code	Plant Pathology Lab	Course type	L	T	P	C	CH
M21SO0Z11		SC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have theoretical knowledge about various plant pathogens and their diseases.

Course Objective:

1. To enumerate the microbial plant pathogens
2. Characterize the plant pathogens for Kochs' postulates
3. Preservation and screening of antagonistic properties of biocontrol agents
4. Mass production of biocontrol agents.

Course Outcome:

By the end of the course the student will be able to

1. Understand the rules of Kochs' postulates.
2. Identification and characterization of disease-causing plant pathogens
3. Maintenance and production of biocontrol agents
4. Preparations and Formulations of fungicides and their usage in disease management.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PS O3
M21SO03Z11	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

1. Demonstration of Koch"s postulates for a fungal plant pathogens
2. Screening for antagonism by dual culture method.
3. Collection and preservation of disease specimens.
4. Extraction of Nematodes from soil and plant material.
5. Transmission of plant viruses and study the virus vector relationship.
6. Evaluation of different fungicides against the fungal pathogens.
7. Preparation of Bordeaux mixture. Burgundy mixture and Chaubattia paste and their practical usage in managing the diseases.
8. Growth observations of different fungal bio agents on different growth media and at different pH regimes.

9. Mass production of *Trichoderma viridae, Paecilomyces lilacinus.*
10. Mass production of Entamopathogenic bio- agents viz, *Metarhizium anisopliae, Verticillium lecanni and Beauveria bassiana.*

References

1. Physiological Plant Pathology; Heitefuss R, Williams PH Springer Verlag, Berlin, New York (1976).
2. Plant Pathology; Mehrotra R S & Aggarwal A 2nd Ed. Oxford &IBH, New Delhi (2003).
3. Introduction to Principles of Plant Pathology; Singh R S. Oxford & IBH, New Delhi (2002).

Course code	Biomass and energy systems Lab	Course type	L	T	P	C	CH
M21SO03Z12		HC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have theoretical knowledge of microorganisms for the energy production.

Course Objective:

1. To understand the soil water plant relations
2. Analyze the quality of various sources of water samples
3. Determine organic carbon in soil samples
4. Formulations of micro and macro nutrients for plants.

Course Outcome:

1. Explore the soil water plant samples and their physio-chemical parameters
2. Apply the knowledge to evaluate the physiochemical parameters of various sources of water samples.
3. Quantitative and qualitative analysis of macro and micro-nutrients
4. Biostatistical data analysis for report preparations.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PSO 3
M21SO03Z12	CO1	3	3	3	2	1		2	2		3	3	3	2

	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Content:

1. Collection and processing of soil, water and plant samples from field
2. Analyses of sewage water samples
3. Analyses of irrigation water samples for pH, EC, SO42-
4. Analyses of irrigation water samples for SAR & RSC
5. Analyses of soil samples for pH, EC, OC and macronutrients
6. Analyses of soil samples for micronutrients
7. Analyses of plant samples for macronutrients
8. Analyses of plant samples for micronutrients
9. Analyses of organic manures
10. Soil, Water and Plant analyses reports preparation.

Reference:

1. Agricultural Microbiology; Rangaswami G and Bagyaraj, Second edition, PHI Learning (P) Ltd., New Delhi (2002).
2. Soil Microbiology; Robert, L Tate Ist edition, John Wiley and Sons, Inc. New York (1995).

Specialization in Medical and Pharmaceutical Microbiology

Course code	Clinical Microbiology	Course type	L	T	P	C	CH
M21SO03Z13		**SC**	**4**	**0**	**0**	**4**	**4**

Prerequisites/Pre reading for the course:

Students should have basics of diseases caused by various classes of microorganisms.

Course Objective:

1. Gain a knowledge on the bacterial diseases and their importance
2. Acquire the knowledge in virology and diseases caused by viruses

3. Understand the life cycle and reproduction of few important protozoans and fungi.
4. Exploit the cutting-edge technology in handling and analysing the pathogens.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PS O3
M21SO03Z13	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Outcome:

By the end of the course the student will be able to:

1. Learn about reproduction in bacteria, aspects of bacterial growth and pathogenic nature of bacterial infections.
2. Know the properties, pathogenicity and lab diagnosis of pathogenic viruses.
3. Obtain the information of source, life cycle and diagnosis of pathogenic protozoans and fungi.
4. Explore the various techniques in the diagnosis and detection of diseases.

Course Content:

Unit – I Bacteriology **13 h**

Bacteria-morphology, ultrastructure, nomenclature and classification, growth and nutrition of bacteria, bacterial toxins, antibacterial substances used in the infections and drug resistance in bacteria. Pathogenic bacteria-*Staphylococcus, Micrococcus, Streptococcus, Neisseria, Corynebacterium, Clostridium, Vibrios, Campylobacter, Haemophilus, Mycobacterium, Pseudomonas, Spirochaetes, Chlamydiae, Rickettsiae, Mycoplasma* & *Ureaplasma*.

Unit – II Virology **13 h**

Characteristics and classification of viruses, morphology, virus structure, viral replication, pathogenicity of viruses, bacteriophages, pox viruses, herpes viruses, arboviruses, orthomyxovirus,

paramyxoviruses, enteroviruses-polio & other enteric viruses, hepatitis viruses, rabies viruses and HIV. Culturing of viruses in embryonated eggs, experimental animals and cell culture. Identification of viruses-physical and chemical methods of assays (protein nuclei acid, radioactivity traces, electrons microscopy and plaque method).

Unit – III Parasitology and Mycology **13 h**

Protozoan parasites of medical importance-Entamoeba, *Giardia, Trichomonas, Leishmania, Trypanosoma, Plasmodium, Toxoplasma* and *Pneumocystis*. Medically important helminths belonging to *Cestodes, Trematodes and Nematodes. Cestodes-Diphyllobothrium, Taenia, Echinnococcu* and *Hymenolepis*, Nematodes-*Trichuris, Trichinella, Strongyloides, Ancylostoma, Ascaris, Enterobius* and *Filarial* worms. The morphology and reproduction in fungi, classification of fungi, opportunistic fungi, superficial mycotic infections, subcutaneous mycoses and systemic infections. Laboratory diagnosis of fungal infections and immunity in fungal diseases.

Unit – IV Immunotechnology **13 h**

Brief about the immune system, innate and adaptive immunity, Antigen and antibodies. Complementation system, Antigens and Antibody reactions in vitro Serological testing - agglutination, complement fixation, ELISA, western blotting, immunodiffusion, immunoelectrophoresis, immunofluorescence, immunoprecipitation, radioimmunoassay (RIA), Flow cytometry and serotyping. Molecular diagnostics – molecular basics of diagnostics, Hybridisation technique and PCR based techniques.

Reference books

1. Medical Microbiology; Brooks, G.F., Butel, J.S., and Ornston, L.N. editors Jawetz, Melnick & Adelberg's 20th ed, Stamford, Conn, Appleton & Lange (1995).
2. Diagnostic microbiology; Forbes, A.B., Sahm, D.F. and Weissfeld, A.S. X Edn. Mosby publishers. New York (2001).
3. Principles and practice of infectious diseases; Mandell, G.L., Bennett, J.E., and Dolin, R 4th ed New York, Churchill Livingstone (1995).
4. The biologic and clinical basis of infectious disease; Raven. Shulman, S.T. Phair, J.P, and Sommers, H.M 4th ed, Philadelphia (1992).
5. Clinical Microbiology. W.B.Saunders. Stokes, J., Ridway, G.L. and Wren, M.W.D., 7th Edn. Edward Arnold – a division of hodder and Stoughton (2013).
6. Immunology; Ian Tizard R 9th edition, Elsevier publisher (2013).
7. Molecular Therapeutics: 21st century medicine; Pamela Greenwell, Michelle McCulley, 1st Edition, Springer (2008).

8. Molecular Diagnostics; George Patrinos, Wilhelm Ansorage, 1st Edition, Academic Press (2005).

Course code	Medical Biochemistry	Course type	L	T	P	C	CH
M21SO03Z14		SC	4	0	0	4	4

Prerequisites/Pre reading for the course:

Students should have basic knowledge of human physiology and biochemical relations.

Course Objective:

1. Acquire the knowledge of source, functions and classification of carbohydrates and lipids.
2. Better understanding of the source, properties and functions associated with the proteins and its clinical relevance.
3. Inculcate the important understanding of disorders associated with the diet and lifestyle.
4. Explore the knowledge of enzymes and their applications in diagnostic purpose.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PS O3
M21SO03Z14	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Outcome:

By the end of the course the student will be able to:

1. Explore the important functions and disorders associated with carbohydrates and lipids.
2. Acquaint the sources, functions and malnutrition associated with proteins.
3. Depth understanding of healthy diet and role of various sectors in health aspects.
4. Apply and analyze the enzymes in the medical applications and diagnostic purpose.

Course Content:

Unit – I Carbohydrates and Lipids **13 h**

Chemistry of carbohydrates and lipids-classifications, properties, classifications. fatty acids types and uses, glycerides, phospholipids, glycolipids, eicosanoids, steroids, cholesterol and lipoproteins.

digestion and absorption of carbohydrates, proteins, fats and nucleic acids. physiology and biochemistry of respiration. Detoxification mechanisms generally taking place in human body and body fluids.

Unit – II Proteins and clinical relevance 13 h

Amino acids-classification based on structure, requirement, metabolic fate, solubility, physical properties and chemical properties of amino acids. Proteins-definition, structure, classification and functions. Gout and genetic defects in urate metabolism. Methods employed usually in protein quality evaluation. Nutritional experiments commonly done on laboratory animals. Recent advance techniques used in human nutrition studies

Unit – III Nutrition and Public Health 13 h

Human dietary requirements and deficiency diseases, BMR and BMI, daily nutritional requirements-macro and micronutrients. Malnutrition of proteins and energy, symptoms and diagnosis of nutritional deficiencies-Anaemia, blindness, beriberi, pellagra. Diet, calculation of balanced diet, disorders of protein energy malnutrition. Health determinants and standards, relevance to social aspects, future challenges in public health. Role of public, private and NGO in Health sector.

Unit – IV Principles of diagnostic enzymology 13 h

Introduction to enzymes and nomenclature. Enzyme kinetics and inhibitors, chromogens in brief. Medically important enzymes and factors affecting enzyme levels in blood. Principle, assay and clinical significance of transaminases, creatine kinase, lactate dehydrogenase, phosphatases, isocitrate dehydrogenase, amylase, lipase, trypsin, chymotrypsin, choline esterase, glutamate dehydrogenase, glucose-6-phosphate dehydrogenase and ceruloplasmin.

Reference books

1. Principles of Biochemistry; Lehninger and D L Nelson, 6th edition, Macmillan Publications (2012).
2. Biochemistry; Jeremy M Berg, John L Toymoczko and Lubert Stryer, 8th Edition Macmillan Publications (2015).
3. Physical Biology of the Cell; Rob Phillips, Jane Kondev, Julie Theriot, Hernan Garcia, 2nd edition, Garland Publishers (2012).
4. Biochemistry; Voet, D. and Voet, J.G. 3rd edition; John Wiley and sons (2009).
5. Biochemistry; Garrett and Charles M. Grisham, 6th edition, Mary Finch publisher (2013).
6. Biochemistry and Molecular Biology; Elliot, 4th edition, Oxford University Press (2009).

7. Health & Wellness; Gordon Edlin and Eric Golanty, 10th Edn Jones & Barlett Publisher (2010).
8. Introduction to Public Health; Mary-Jane Schneider 4th Edn Jones & Barlett (2014).

Course code	Pharmaceutical Microbiology	Course type	L	T	P	C	CH
M21SO03Z15		SC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have basic knowledge of microbes and their importance in pharma industries.

Course Objective:

1. Acquire the knowledge about antibiotics and their various usage in industries
2. Better understanding of the molecular mechanism of action of various antibiotics
3. Exploit the technology for the microbial production of antibiotics and their spoilage
4. Establish the quality assurance and data management in the pharma industries.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PS O3
M21SO03Z15	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Outcome:

By the end of the course the student will be able to:

1. In-depth understanding about various types of antibiotics and their applications
2. Identify the molecular mechanistic insight of antibiotics actions
3. Utilize the technological advancement for production of antibiotics
4. Explore the quality assurance and quality control aspects involved in industries.

Course Content:

Unit – I Antibiotics and synthetic antimicrobial agents **13 h**

Antibiotics and synthetic antimicrobial agents - Aminoglycosides, β lactams, tetracyclines, ansamycins, macrolid antibiotics. Antifungal antibiotics, antitumor substances. Peptide antibiotics, chloramphenicol, sulphonamides and quinolinone antimicrobial agents. Chemical disinfectants, antiseptics and preservatives.

Unit – II Mechanism of action of antibiotics 13 h

Mechanism of action of antibiotics. Molecular principles of drug targeting. Drug delivery system in gene therapy Bacterial resistance to antibiotics. Mode of action of bacterial killing by quinolinones. Bacterial resistance to quionolinones. Mode of action of non – antibiotic antimicrobial agents.

Unit – III Microbial production and spoilage of pharmaceutical products 13 h

Microbial contamination and spoilage of pharmaceutical products (sterile injectables, non injectibles, ophthalmic preparations and implants) and their sterilization. Manufacturing procedures and in process control of pharmaceuticals. Other pharmaceuticals produced by microbial fermentations (streptokinase, streptodornase). New vaccine technology, DNA and RNA vaccines, synthetic peptide vaccines and multivalent subunit vaccines.

Unit – IV Quality Assurance and Validation 13 h

Good Manufacturing Practices (GMP) and Good Laboratory Practices (GLP) in pharmaceutical industry. Regulatory aspects of quality control. Quality assurance and quality management in pharmaceuticals ISO, WHO and US certification. Sterilization control and sterility testing-chemical and biological indicators. Design and layout of sterile product manufacturing unit. Designing of Microbiology laboratory and safety in microbiology laboratory. Risk management and Risk Mitigations.

Reference books

1. Pharmacology and Pharmacotherapeutics; R.S. Satoskar and Nirmala N Rege, Raakhi K. Tripathi, S.D. Bhandarkar, 2nd Edition, Popular Prakashan Publishers (2010).
2. Microbiology: Principles & Explorations; JG Black, 6th edn, John Wiley & Sons (2010).
3. Free, Hydrometallurgy: Fundamentals and Applications; Michael L, 1st edition, Wiley Publications (2013).
4. Biomining: Theory, Microbes and Industrial Processes (Biotechnology Intelligence Unit); Douglas E. Rawlings, 1st Edition, Springer publications (2012).

5. The Science and practice of Pharmacy; 20th edition - David B. Troy, Paul Beringer Lippincott Williams and Wilkins (2010).

Course code		Course type	L	T	P	C	CH
M21SO03Z16	Clinical Microbiology LAB	SC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have theoretical knowledge about physiological functions of host during disease conditions.

Course Objective:

1. In-depth understanding the establishment setup of the clinical microbiology laboratory and good lab practices.
2. Hands on experiences in handling and preservations of the laboratory specimens.
3. Exposure to the various techniques in molecular diagnostic protocols.
4. Explore the basics of the molecular diagnostic laboratory.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PS O3
M21SO03Z16	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Outcome:

By the end of the course the student will be able to:

1. Better understanding of the clinical microbiology laboratory setup.
2. Exploit the handling of the clinical specimens for diagnostics.
3. Explore the techniques involved in the pre-processing of molecular diagnostics.
4. Exploit the various advanced molecular techniques for diagnostics and research.

Course Content:

1. Laboratory safety: Fire, chemical, radiation, handling of biological specimens, waste, Disposal regulations and workplace hazardous.
2. Specimen collection, identification, transport, delivery and preservation.
3. ELISA technique-direct.
4. Indirect ELISA and analyses.
5. PCR for virus detection.
6. RT-PCR quantitation of viruses.
7. Separation of serum from blood and preparation for further testing.
8. Isolation of DNA followed by analysis through agarose gel electrophoresis.
9. Isolation of RNA followed by analysis through formaldehyde agarose gel electrophoresis.
10. Quality and quantitative analysis of DNA and RNA by spectrophotometer.

Reference

1. Molecular Diagnostics: Fundamentals, Methods and Clinical Applications; Lela Buchingham and Maribeth L Flawsm, 1st Edition, F A Davis Company, Philadelphia, USA (2007).
2. Immunology; Kuby J, Judy Owen, Jenni Punt, Sharon Stranford; 7th Edition. W.H. Freeman and Company (2013).
3. Essential Immunology: Peter J. Delves, Seamus J. Martin, Dennis R. Burton, Ivan M. Roitt, 12th edition, ELBS, Blackwell Scientific Publishers, London (2011).

Course code	Medical Biochemistry LAB	Course type	L	T	P	C	CH
M21SO03Z17		HC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have theoretical knowledge of biochemical aspects in human abnormal conditions.

Course Objective:

1. Understand the various protein extraction and analysis techniques
2. Establishment protocols and setup of Medical Biochemistry lab.

3. Hands-on experience in analysis of the cells and sub-cellular parts
4. Explore various molecular mechanism of microRNA studies.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PS O1	PS O2	PS O3
M21SO03Z17	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Outcome:

By the end of the course the student will be able to:

1. Better understanding of the biochemical methods in protein extraction and analysis.
2. Hand-on experience in advance methods of flowcytometry and microRNA.
3. Standardize the protocols for the medical biochemistry lab.
4. Exploit the molecular and biochemical analysis methods in industries and research.

Course Content:

1. Tissue homogenization and cell disruption-Cell fractionation methods by ultra and gradient centrifugation
2. Protein extraction and column purification from the samples and its spectrophotometer analysis.
3. Flowcytometric analysis of blood cells.
4. Gelatin zymographic analysis.
5. SDS PAGE for isozymes detection.
6. Western blotting analysis for the gene products-biomarker analysis.
7. Preparation of immunohistochemistry samples and confocal/fluorescent microscopy.
8. Preparation of samples for electron microscopy and demonstration of imaging.
9. Estimation of LDH and phosphatase levels in blood with clinical significance.
10. Isolation and identification of miRNA.

Reference books

1. Analytical techniques in Biochemistry and molecular Biology; Katoch, Rajan, Springer (2011).

2. Basic Methods for the Biochemical Lab; Martin Holtzhauer, Springer (2007).
3. Principles and Techniques of Biochemistry and Molecular Biology; Keith Wilson and John Walker 7th edition., Cambridge University Press (2010).
4. Experiments in Applied Microbiology; Samuel Singer, Academic Press (2001).

Course code	Pharmaceutical Microbiology LAB	Course type	L	T	P	C	CH
M21SO03Z18		HC	0	0	4	2	4

Prerequisites/Pre reading for the course:

Students should have theoretical knowledge of microbial quality control in pharmaceutical industries.

Course Objective:

1. Understand the microbiological methods for determination of antibiotics.
2. Practical explore in the antimicrobial sensitivity assay for antibiotics.
3. Aid in conducting the toxicity studies in the cell culture.
4. Determine the microbial spoilage and load in the pharmaceutical products.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PSO 3
M21SO03Z18	CO1	3	3	3	2	1		2	2		3	3	3	2
	CO2	3	2	3	1	1		2	2		3	3	3	2
	CO3	3	2	3	1	2		2	2		3	3	3	2
	CO4	3	3	3	2	1		2	2		3	3	3	2

Course Outcome:

By the end of the course the student will be able to:

1. Determine the microbial methods in the antibiotic's studies.
2. Exploit the antimicrobial sensitivity assay for quality analysis and research.
3. Assist and analyze the toxicity studies in the cell cultures.

4. Analyze the various microbial spoilage in the pharmaceutical products.

Course Content:

1. Microbiological methods for the determination of griseofulvin/streptomycin.
2. Bioassay of chloramphenicol by plate assay method.
3. Antimicrobial susceptibility testing by broth dilution method.
4. To determine MIC, LD50 of Beta-lactam/aminoglycoside/ tetracycline.
5. Sterility testing by *Bacillus stearothermophilus*
6. Sampling of pharmaceuticals for microbial contamination and load (syrups, suspensions, creams and ointments, ophthalmic preparations).
7. Separation of mononuclear cells by Ficoll-Hypaque and cell viability by MTT assay.
8. Identification of various types of blood cells by blood smear and microscopy.
9. Isolation of peripheral blood cells and viability testing by live-dead staining method.
10. Counting and viability assessment of animal cells by direct microscopic methods.

Reference

1. Microbiological methods; Collins, C.H., Tatrica M. Lyne and Grange, J.M. 8th edition. Hodder Arnold publishers (2004).
2. Biopharmaceuticals: Biochemistry and Biotechnology; Gary Walsh, 2nd Edition, John Wiley & Sons, Wiley Publications (2013).
3. Biotechnology & Biopharmaceuticals Transforming Proteins and Genes into Drugs; Rodney J. Y. Ho 2nd Edn Wiley-Blackwell (2013).

FOURTH SEMESTER

Course Code	Course Title	Course Type	L	T	P	C	Hrs./ Wk.
M21SO0401	**MAJOR PROJECT AND DISSERTATION**	HC	0	0	10	10	10

Course Objective:

To carry out the research under the guidance of supervisor and in the process learn the techniques of research.

Course Outcome:

On successful completion of the project, the student shall be able to:

1. Apply fundamental and disciplinary concepts and methods in ways appropriate to their principal areas of study.
2. Demonstrate the skill sets acquired and employ the knowledge of current information in the domain.
3. Apply technological tools and techniques specific to the professional field of study.
4. Acquire real time exposure to the systematic execution of research components and methodology.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SO0401	CO1	2	3		3							3	2	2
	CO2	2	2	3	3	3	3					2	3	2
	CO3	2	2	3	3	3	3		3	3	3	3	3	3
	CO4	3	3	3	3	3	3	3	3	3	3	3	3	3

Course Content:

Each student individually select the topic of research preferably from any area of core domains of microbiology and work under the guidance of allocated faculty member. The project shall preferably be application oriented or industry need based that could be useful to the society. In case of industry need based project or R & D project the student may opt co-supervisor from the concerned industry / research institution as the case may be. The student will have to make a preliminary survey of research done in broad area of his/her area of interest and decide on the topic in consultation with his/her supervisor(s). The project work floated should be completed within 6-months and project report has to be submitted within the stipulated date by the University/ within 6-months whichever is earlier. The student has to meet the concerned supervisor(s) frequently to seek guidance and also to produce the progress of the work being carried out. The student should also submit progress report during 5th week and 10th week of the beginning of the semester and final draft report with findings by 14th week. After the completion of the project the student shall submit project report in the form of dissertation on a specified date by the School.

Course Code	Course Title	Course Type	L	T	P	C	Hrs./ Wk.
M21SOON01	MOOC / SWAYAM 1	-	-	-	-	2	-
M21SOON02	MOOC / SWAYAM 2	-	-	-	-	2	-

Course Objective:

To carry out the e-learning from the SWAYAM and NPTEL in the subject domains for enhance the knowledge of related areas from the reputed institutes in India.

Course Outcome:

1. Enhance the conceptual knowledge and ensure academic achievement in the subject domain offered through the e learning.
2. Gain additional expertise gained in the process of e-learning.
3. Acquire intellectual skills such as domain-specific and generic abilities involved in reasoning, comprehension and thinking abilities.
4. Inculcate the problem-solving, and decision-making skills related to the subject domain.

Mapping of Course Outcomes with Program Outcomes

Course Code	POS / COS	PO1	PO2	PO3	PO 4	PO5	PO6	PO7	PO8	PO9	PO10	PSO 1	PS O2	PS O3
M21SOON01 M21SOON02	CO1	2	2	3			3					1	1	3
	CO2	3	2									1	1	3
	CO3	3	3								3	1	1	3
	CO4	3	3								3	1	1	3

Course Content:

MOOC / SWAYAM Online Courses: Globally, MOOC (Massive Open Online Course) platforms are gaining much popularity. Considering the popularity and relevance of MOOCs, Government of India has also launched an indigenous platform, SWAYAM. SWAYAM (Study Webs of Active Learning for Young Aspiring Minds) is basically an integrated MOOCs platform for distance education that is aimed at offering all the courses from school level (Class IX) to post-graduation

level. The platform has been developed collaboratively by MHRD (Ministry of Human Resource Development) and AICTE (All India Council for Technical Education) with the help of Microsoft and is capable of hosting 2,000 courses.

Students are required to complete courses like technical skills, placement related courses, Open electives and any such value addition or specialized courses through online platforms like SWAYAM/NPTEL/Any other reputed online education aggregator. Students are required to choose the courses on the advice of their course coordinator/Director and required to submit the course completion certificate along with percentage of marks/grade scored in the assessment conducted by the online education aggregator. If the online education aggregator has issued a certificate along with the grade or marks scored to students, such courses will be considered for SGPA calculations, in case the aggregator has issued only a certificate and not marks scored, then such courses will be graded through an examination by concerned School, in case, if grading is not possible, students will be given a pass grade and award the credit and the credits will not be considered for SGPA calculations. The Online/MOOCs courses will not have continuous internal assessment component.

CAREER OPPORTUNITIES

Having a degree will open doors to the world of opportunities for you. But Employers are looking for much more than just a degree. They want graduates who stand out from the crowd and exhibit real life skills that can be applied to their organizations. Examples of such popular skills employers look for include:

1. Willingness to learn
2. Self-motivation
3. Team work
4. Communication skills and application of these skills to real scenarios
5. Requirement of gathering, design and analysis, development and testing skills
6. Analytical and Technical skills
7. Computer skills
8. Internet searching skills
9. Information consolidation and presentation skills
10. Role play
11. Group discussion, and so on

REVA University therefore, has given utmost importance to develop these skills through variety of training programs and such other activities that induce the said skills among all students. A full-fledged Career Counselling and Placement division, namely Career Development Centre (CDC) headed by well experienced senior Professor and Dean and supported by dynamic trainers, counsellors and placement officers and other efficient supportive team does handle all aspects of Internships and placements for the students of REVA University. The prime objective of the CDC is to liaison between REVA graduating students and industries by providing a common platform where the prospective employer companies can identify suitable candidates for placement in their respective organization. The CDC organizes pre-placement training by professionals and also arranges expert talks to our students. It facilitates students to career guidance and improve their employability. In addition, CDC forms teams to perform mock interviews. It makes you to enjoy working with such teams and learn many things apart from working together in a team. It also makes you to participate in various student clubs which helps in developing team culture, variety of job skills and overall personality.

The need of the hour in the field of Microbiology and Life Sciences is not only knowledge in the subject, but also the skills to do the job proficiently, team spirit and a flavour of innovation. This

kept in focus, the CDC has designed the training process, which will commence from second semester along with the curriculum. Special coaching in personality development, career building, English proficiency, reasoning, puzzles, and communication skills to every student of REVA University is given with utmost care. The process involves continuous training and monitoring the students to develop their soft skills including interpersonal skills that will fetch them a job of repute in the area of his / her interest and march forward to make better career. The School of Chemical and Biological sciences also has emphasised subject based skill training through lab practice, internship, project work, industry interaction and many such skilling techniques. The students during their day to day studies are made to practice these skill techniques as these are inbuilt in the course curriculum. Concerned teachers also continuously guide and monitor the progress of students.

The University has also established University-Industry Interaction and Skill Development Centre headed by a Senior Professor & Director to facilitate skill related training to REVA students and other unemployed students around REVA campus. The centre conducts variety of skill development programs to students to suite to their career opportunities. Through this skill development centre the students shall compulsorily complete at least two skill / certification based programs before the completion of their degree. The University has collaborations with Industries, Corporate training organizations, research institutions and Government agencies like NSDC (National Skill Development Corporation) to conduct certification programs. REVA University has been recognised as a Centre of Skill Development and Training by NSDC (National Skill Development Corporation) under Pradhan Mantri Kaushal VikasYojana. The University has also signed MOU's with Multi-National Companies, research institutions, and universities abroad to facilitate greater opportunities of employability, students' exchange programs for higher learning and for conducting certification programs.